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(54) Title: USE OF ANTIBODIES TO THE GAMMA 2 CHAIN OF LALLMININ 5 TO INHIBIT TUMOR GROWTH AND METASTASIS

(57) Abstract: The present invention provides a methods and compositions for inhibiting tumor growth and/or metastasis involving the administering to a subject with a laminin 5-secreting tumor of an amount effective to inhibit tumor growth and/or metastasis of an antibody that binds to one or more epitopes of the laminin 5 γ 2 chain.

WO 2004/039401 A2

Use of antibodies to the gamma 2 chain of laminin 5 to inhibit tumor growth and metastasis

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CROSS REFERENCE

This application claims priority to U.S. Provisional Patent Application 60/422,009 filed October 29, 2002.

10 BACKGROUND OF THE INVENTION

Laminins are basement membrane glycoproteins with diverse biological functions including cell adhesion, proliferation, migration and differentiation. Thus far, 11 genetically distinct chains forming at least 12 laminin isoforms have been characterized. Every member of this growing protein family has a heterotrimeric chain composition of α , β , and γ chains that are
15 formed through an intracellular self-assembly mechanism.

Laminin-5 is a specific component of epithelial basement membranes with the chain composition $\alpha 3\beta 3\gamma 2$ (Kallunki, et al., J. Cell Biol. 119: 679-93, 1992). The $\gamma 2$ chain has a mass of ≈ 130 kd and is thus smaller than the "classical" ≈ 200 kd $\beta 1$ and $\gamma 1$ light chains of laminin 1. Expression of laminin 5 chains is often up-regulated in epithelial cancers, such as squamous cell
20 carcinomas and gastric carcinomas, but not in mesenchymally derived cancers (Larjava, et al., J. Clin. Invest. 92: 1425-35, 1993) (Pyke, et al., Am. J. Pathol. 145: 782-91, 1994) (Pyke, et al., Cancer Res. 55: 4132-9, 1995) (Tani, et al., Am. J. Pathol. 149: 781-93, 1996) (Orian-Rousseau, et al., J. Cell. Sci. 111: 19932004, 1998) (Sordat, et al., J. Pathol. 185: 44-52, 1998). However, down-regulation has been reported in epithelial prostate and breast carcinomas (Hao, J., Yang,
25 Am. J. Pathol. 149: 1341-9, 1996) (Martin, et al., Mol. Med. 4: 602-613, 1998). In colon adenocarcinomas, both gene and protein expression of the $\gamma 2$ chain seem to be a characteristic of cancer cells with a budding phenotype (Larjava, et al., J. Clin. Invest. 92: 1425-35, 1993) (Pyke, et al., Am. J. Pathol. 145: 782-91, 1994) (Pyke, et al., Cancer Res. 55: 4132-9, 1995). Tumor cell budding in colorectal carcinoma has also been associated with the presence of intracellular
30 laminin-5 (Sordat, et al., J. Pathol. 185: 44-52, 1998).

The $\gamma 2$ chain of laminin-5 has also been shown to be strongly expressed in malignant cells located at the invasion front of several human carcinomas, as determined by *in situ* hybridization and immunohistochemical staining (Pyke, C., Romer, J., Kallunki, P., Lund, L.R., Ralfkiaer, E., Dano, K. & Tryggvason, K. (1994) *Am. J. Pathol.* 145: 782-791; Pyke, C., Salo, S., Ralfkiaer, E., Romer, J., Dano, K. & Tryggvason, K. (1995) *Cancer Res.* 55: 4132-4139). However, no studies have shown that antibodies to the $\gamma 2$ chain of laminin 5 can be used to inhibit tumor cell growth.

SUMMARY OF THE INVENTION

The present invention provides antibodies, compositions and methods for inhibiting tumor growth and/or metastasis. In one aspect, the present invention provides antibodies that bind to one or more epitopes of domain III of the human laminin 5 $\gamma 2$ chain (SEQ ID NOS: 2 and 4).

In another aspect, the present invention provides a method for inhibiting tumor growth and/or metastasis comprising administering to a subject with a laminin 5-secreting tumor an amount effective to inhibit tumor growth and/or metastasis of an antibody that binds to one or more epitopes of the laminin 5 $\gamma 2$ chain. In one embodiment, the antibody binds to one or more epitopes of domain III of the laminin 5 $\gamma 2$ chain.

In a further aspect, the present invention provides a pharmaceutical composition comprising an antibody that binds to the laminin 5 $\gamma 2$ chain and one or more further anti-tumor agents. In various embodiments of this aspect, the antibody is selective for one or more epitopes in domain III of the laminin 5 $\gamma 2$ chain, and/or the further anti-tumor agent is a chemotherapeutic.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGURE 1 shows efficiency of human laminin-5 and recombinant human laminin $\gamma 2$ chain for attachment of HaCat keratinocytes and KLN205 squamous carcinoma cells *in vitro*. The attachment efficiency was compared with the efficiency with which the cells bound to laminin-1. Substrate concentrations (10 $\mu\text{g/ml}$) providing maximum attachment to laminin-1 and laminin-5 were used. The results are presented as means \pm SD calculated from at least four duplicate series; the values for laminin-1 were given the arbitrary value of 100%.

FIGURES 2A-B show the effects of polyclonal $\gamma 2$ chain antibodies on the migration of KLN205 squamous carcinoma cells in Boyden and Transwell chamber assays of migration.

FIGURE 3 shows tumor growth inhibition using Mab 5D5 and CPT-11 on day 31 in the HT29-e28 cell line.

5 FIGURES 4A-E show tumor growth curves for individual mice in the HT29-e28 study.

DETAILED DESCRIPTION OF THE INVENTION

10 In one aspect, the present invention provides a method for inhibiting tumor growth and/or metastasis comprising administering to a subject with a laminin 5-secreting tumor an amount effective to inhibit tumor growth and/or metastasis of an antibody that binds to one or more epitopes of the laminin 5 $\gamma 2$ chain. In a preferred embodiment, the subject is a mammal; in a more preferred embodiment, the subject is human.

15 As used herein, the term "inhibiting tumor growth" means to reduce the amount of tumor growth that would occur in the absence of treatment, and includes decrease in tumor size and/or decrease in the rate of tumor growth.

As used herein, the term "inhibiting tumor metastasis" means to reduce the amount of tumor metastasis that would occur in the absence of treatment, and includes decrease in the number and/or size of metastases.

20 As used herein, the term "laminin-5 secreting tumor" means a tumor that expresses detectable amounts of laminin 5. Such tumors include, but are not limited to, carcinomas. Such carcinomas include, but are not limited to squamous cell carcinomas (including but not limited to squamous cell carcinoma of skin, cervix, and vulva), gastric carcinomas, colon adenocarcinomas, colorectal carcinomas, and cervical carcinomas.

25 As used herein, the term "laminin 5 $\gamma 2$ chain" preferably refers to the human laminin 5 $\gamma 2$ chain, with protein sequences comprising the amino acid sequence of SEQ ID NO:2 or SEQ ID NO:4, and derivatives thereof.

As used herein, the term "epitope" refers to a specific site within the protein that is bound by the antibody, which includes both linear and non-linear epitopes.

30 In a preferred embodiment, the antibody binds to one or more epitopes of domain III of the laminin 5 $\gamma 2$ chain. As used herein, the term "domain III of the laminin 5 $\gamma 2$ chain" refers to

a 177 amino acid region of SEQ ID NO:2 between residues 391 and 567 (Kallunki et al., J. Cell Biol. 119:679-693 (1992)), which is presented herein as SEQ ID NO:8. In a further preferred embodiment, the antibody binds to one or more epitopes within domain III that are contained within the amino acid sequence of SEQ ID NO:6 and does not bind to epitopes within domain
5 III that are within the amino acid sequence of SEQ ID NOS: 9 and 10.

The antibody can be a polyclonal antibody or a monoclonal antibody, but preferably is a monoclonal antibody. For use in humans, humanized monoclonal antibodies are especially preferred.

In a further embodiment, the methods of the invention further comprises treating the
10 subject with chemotherapy and/or radiation therapy, whereby the use of the antibody permits a reduction in the chemotherapy and/or radiation dosage necessary to inhibit tumor growth and/or metastasis. Any reduction in chemotherapeutic or radiation dosage benefits the patient by resulting in fewer and decreased side effects relative to standard chemotherapy and/or radiation therapy treatment.

In this embodiment, the antibody may be administered prior to, at the time of, or shortly
15 after a given round of treatment with chemotherapeutic and/or radiation therapy. In a preferred embodiment, the antibody is administered prior to or simultaneously with a given round of chemotherapy and/or radiation therapy. In a most preferred embodiment, the antibody is administered prior to or simultaneously with each round of chemotherapy and/or radiation therapy.
20 The exact timing of antibody administration will be determined by an attending physician based on a number of factors, but the antibody is generally administered between 24 hours before a given round of chemotherapy and/or radiation therapy and simultaneously with a given round of chemotherapy and/or radiation therapy.

The methods of the invention are appropriate for use with chemotherapy using one or more
25 cytotoxic agent (ie: chemotherapeutic), including, but not limited to, cyclophosphamide, taxol, 5-fluorouracil, adriamycin, cisplatin, methotrexate, cytosine arabinoside, mitomycin C, prednisone, vindesine, carbaplatin, and vincristine. The cytotoxic agent can also be an antiviral compound which is capable of destroying proliferating cells. For a general discussion of cytotoxic agents used in chemotherapy, see Sathe, M. et al., Cancer Chemotherapeutic Agents: Handbook of
30 Clinical Data (1978), hereby incorporated by reference.

The methods of the invention are also particularly suitable for those patients in need of repeated or high doses of chemotherapy and/or radiation therapy.

In practicing the invention, the amount or dosage range of antibody employed is one that effectively inhibits tumor growth and/or metastasis. The actual dosage range is based on a variety of factors, including the age, weight, sex, medical condition of the individual, the severity of the condition, and the route of administration. An inhibiting amount of antibody that can be employed ranges generally between 0.01 $\mu\text{g/kg}$ body weight and 15 mg/kg body weight, preferably ranging between 0.05 $\mu\text{g/kg}$ and 10 mg/kg body weight, more preferably between 1 $\mu\text{g/kg}$ and 10 mg/kg body weight, and even more preferably between about 10 $\mu\text{g/kg}$ and 5 mg/kg body weight.

The antibody may be administered by any suitable route, but is preferably administered parenterally in dosage unit formulations containing conventional pharmaceutically acceptable carriers, adjuvants, and vehicles. The term "parenteral" as used herein includes, subcutaneous, intravenous, intraarterial, intramuscular, intrasternal, intratendinous, intraspinal, intracranial, intrathoracic, infusion techniques or intraperitoneally. In preferred embodiments, antibody is administered intravenously or subcutaneously.

The antibody may be made up in a solid form (including granules, powders or suppositories) or in a liquid form (e.g., solutions, suspensions, or emulsions). Antibody may be applied in a variety of solutions. Suitable solutions for use in accordance with the invention are sterile, dissolve sufficient amounts of the antibody, and are not harmful for the proposed application.

The antibody may be subjected to conventional pharmaceutical operations such as sterilization and/or may contain conventional adjuvants, such as preservatives, stabilizers, wetting agents, emulsifiers, buffers etc.

For administration, the antibody is ordinarily combined with one or more adjuvants appropriate for the indicated route of administration. The compounds may be admixed with lactose, sucrose, starch powder, cellulose esters of alkanolic acids, stearic acid, talc, magnesium stearate, magnesium oxide, sodium and calcium salts of phosphoric and sulphuric acids, acacia, gelatin, sodium alginate, polyvinylpyrrolidone, and/or polyvinyl alcohol, and tableted or encapsulated for conventional administration. Alternatively, the antibody may be dissolved in saline, water, polyethylene glycol, propylene glycol, carboxymethyl cellulose colloidal solutions, ethanol, corn oil, peanut oil, cottonseed oil, sesame oil, tragacanth gum, and/or various buffers.

Other adjuvants and modes of administration are well known in the pharmaceutical art. The carrier or diluent may include time delay material, such as glyceryl monostearate or glyceryl distearate alone or with a wax, or other materials well known in the art.

5 In another aspect, the present invention provides isolated antibodies that bind to one or more epitopes of domain III of the laminin 5 γ 2 chain, hybridoma cells that produce isolated monoclonal antibodies, and pharmaceutical compositions comprising such monoclonals. In a further preferred embodiment, the isolated antibody binds to one or more epitopes within the amino acid sequence of **SEQ ID NO:6** and does not bind to epitopes within the amino acid sequence of **SEQ ID NOS: 9 and 10**. The isolated antibody can be a polyclonal antibody or a
10 monoclonal antibody, but preferably is a monoclonal antibody. In a further embodiment, the isolated antibodies are humanized. In a further embodiment, the isolated antibody is prepared as a pharmaceutical composition, combined with one or more appropriate pharmaceutical carriers, as described above.

These isolated antibodies are useful in all of the methods of the invention, as well as in
15 diagnostic use for detecting the presence of invasive cells in a tissue sample. In a preferred embodiment, diagnostic use of the isolated antibodies of the invention comprises contacting a tumor tissue with one or more isolated antibodies to form an immunocomplex, and detecting formation of the immunocomplex, wherein the formation of the immunocomplex correlates with the presence of invasive cells in the tissue. The contacting can be performed *in vivo*, using
20 labeled isolated antibodies and standard imaging techniques, or can be performed *in vitro* on tissue samples.

In a preferred embodiment, the tissue is a tumor tissue. In a further preferred embodiment, the tumor tissue is a laminin 5 secreting tumor tissue. More preferably, the tumor tissue is a carcinoma, including but are not limited to squamous cell carcinomas (including but
25 not limited to squamous cell carcinoma of skin, cervix, and vulva), gastric carcinomas, colon adenocarcinomas, colorectal carcinomas, and cervical carcinomas.

In a further preferred embodiment of this aspect of the invention, the isolated monoclonal antibodies are of the IgG isotype. In a further preferred embodiment, the isolated monoclonal antibodies are selected from the group consisting of those designated herein as 4G1, 5D5 and
30 6C12, and the hybridomas expressing these monoclonals, which are deposited with the American Type Tissue Collection as ATCC accession numbers ----, ----, and ----. A more detailed

description of the production of these particular hybridomas and monoclonal antibodies, and their use, is provided below.

The additional components of pharmaceutical compositions comprising one or more of these isolated antibodies are as described above.

5 Antibodies can be made by well-known methods, such as described in Harlow and Lane, Antibodies; A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., (1988). In one example, pre-immune serum is collected prior to the first immunization. A peptide portion of the amino acid sequence of a laminin 5 γ 2 chain polypeptide, together with an appropriate adjuvant, is injected into an animal in an amount and at intervals sufficient to elicit
10 an immune response. Animals are bled at regular intervals, preferably weekly, to determine antibody titer. The animals may or may not receive booster injections following the initial immunization. At about 7 days after each booster immunization, or about weekly after a single immunization, the animals are bled, the serum collected, and aliquots are stored at about -20° C. Polyclonal antibodies against the laminin 5 γ 2 chain polypeptides can then be purified directly by
15 passing serum collected from the animal through a column to which non-antigen-related proteins prepared from the same expression system without the laminin 5 γ 2 chain polypeptides bound.

Monoclonal antibodies can be produced by obtaining spleen cells from the animal. (See Kohler and Milstein, Nature 256, 495-497 (1975)). In one example, monoclonal antibodies (mAb) of interest are prepared by immunizing inbred mice with a laminin 5 γ 2 chain polypeptide,
20 or portion thereof. The mice are immunized by the IP or SC route in an amount and at intervals sufficient to elicit an immune response. The mice receive an initial immunization on day 0 and are rested for about 3 to about 30 weeks. Immunized mice are given one or more booster immunizations of by the intravenous (IV) route. Lymphocytes from antibody positive mice are obtained by removing spleens from immunized mice by standard procedures known in the art.
25 Hybridoma cells are produced by mixing the splenic lymphocytes with an appropriate fusion partner under conditions that allow formation of stable hybridomas. The antibody producing cells and fusion partner cells are fused in polyethylene glycol at concentrations from about 30% to about 50%. Fused hybridoma cells are selected by growth in hypoxanthine, thymidine and aminopterin supplemented Dulbecco's Modified Eagles Medium (DMEM) by procedures known
30 in the art. Supernatant fluids are collected from growth positive wells and are screened for antibody production by an immunoassay such as solid phase immunoradioassay. Hybridoma

cells from antibody positive wells are cloned by a technique such as the soft agar technique of MacPherson, Soft Agar Techniques, in Tissue Culture Methods and Applications, Kruse and Paterson, Eds., Academic Press, 1973.

To generate such an antibody response, a laminin 5 γ 2 chain polypeptide or portion thereof is typically formulated with a pharmaceutically acceptable carrier for parenteral administration. Such acceptable adjuvants include, but are not limited to, Freund's complete, Freund's incomplete, alum-precipitate, water in oil emulsion containing *Corynebacterium parvum* and tRNA. The formulation of such compositions, including the concentration of the polypeptide and the selection of the vehicle and other components, is within the knowledge of those of skill of the art.

The term antibody as used herein is intended to include antibody fragments thereof which are selectively reactive with the laminin 5 γ 2 chain polypeptides. Antibodies can be fragmented using conventional techniques, and the fragments screened for utility in the same manner as described above for whole antibodies. For example, F(ab')₂ fragments can be generated by treating antibody with pepsin. The resulting F(ab')₂ fragment can be treated to reduce disulfide bridges to produce Fab' fragments.

In another aspect, the present invention provides pharmaceutical compositions comprising an antibody that binds to the laminin 5 γ 2 chain and one or more further anti-tumor agent. In a preferred embodiment of this aspect of the invention, the antibody binds to one or more epitopes in domain III of the laminin 5 γ 2 chain, as described above. In a further preferred embodiment, the isolated antibody binds to one or more epitopes within the amino acid sequence of SEQ ID NO:6 and does not bind to epitopes within the amino acid sequence of SEQ ID NOS: 9 and 10. The antibody can be a polyclonal antibody or a monoclonal antibody, but preferably is a monoclonal antibody. In a further preferred embodiment of this aspect of the invention, the further anti-tumor agent is a chemotherapeutic agent, such as one or more of those described above. The components of the pharmaceutical composition may be pre-mixed together or may be combined at any time prior to administration to a patient in need thereof.

The examples below are meant by way of illustration, and are not meant to be limiting as to the scope of the instant disclosure.

EXAMPLE 1

The following example demonstrates the effect of laminin-5, including the $\gamma 2$ chain of laminin-5, on cell adhesion and cell migration.

5 *Materials and Methods**Cells and Cell Culture*

A mouse squamous cell carcinoma cell line, KLN205 (cat. no. ATCC CRL-1453), was obtained from American Type Culture Collection (Rockville, MD). The cells were maintained as monolayer cultures in Eagle's minimum essential medium (MEM) containing non-essential
10 amino acids and Earle's BSS supplemented with 10% fetal calf serum (FCS). The HaCat human keratinocyte cell line was a kind gift from Dr. Fuzenig (Heidelberg, Germany). The HaCat cells were cultured in Dulbecco's MEM supplemented with 10% FCS. However, when the cells were cultured for the production of laminin-5, the medium was replaced by serum-free medium.

15 *Preparation of Proteins*

Mouse EHS laminin (laminin-1) was obtained from GIBCO BRL. Fibronectin was purified from FCS using a gelatin-Sepharose 4B column (Sigma) as described in Vuento, M. & Vaheri, A. (1979) Biochem. J. 183: 331-337.34 and Gillies, R. J., Didier, N. & Denton, M. (1986) Anal. Biochem. 159: 109-113. Human laminin-5 was immunoaffinity purified from the
20 media of HaCat cells cultured for three days in the absence of serum. Briefly, the medium was first passed through a 5 ml gelatin-Sepharose column (Sigma, St. Louis, MO) to ensure the complete absence of fibronectin from the protein preparation, after which the medium was passed through a 10 ml anti-laminin $\gamma 2$ -Sepharose affinity column in order to bind laminin-5 molecules. Both columns were equilibrated in phosphate-buffered saline. The anti-laminin $\gamma 2$ -
25 Sepharose affinity column was prepared by coupling a Protein A-purified anti- $\gamma 2$ IgG (8 mg/ml) to 10 ml of CNBr-activated Sepharose (Pharmacia, Uppsala, Sweden). The anti- $\gamma 2$ IgG was purified from a rabbit polyclonal antiserum prepared against a GST-fusion protein containing domain III of the $\gamma 2$ chain (Pyke, C., Salo, S., Ralfkiaer, E., Romer, J., Dano, K. & Tryggvason, K. (1995) Cancer Res. 55: 4132-4139). The laminin-5 was eluted from the immunoaffinity
30 column using 50 mM triethanolamine, pH 11.25, 0.1% Triton X-100 and neutralized directly

with 1 M Tris-HCl, pH 7.0. Collected fractions were analyzed by SDS-PAGE and Western blotting using the same polyclonal antibodies as used for the preparation of the affinity column. Fractions containing laminin-5 were pooled and dialyzed against 50 mM Tris-HCl, 0,1 M NaCl, pH 7.4. Some batches of laminin-5 were denatured with 5 M urea and renatured to study the effects of the treatment on adhesion and migration properties.

Generation of Recombinant Baculovirus and Expression of Recombinant Laminin 2 Chain

The $\gamma 2$ chain of laminin-5 was expressed as recombinant protein using the baculovirus system and purified for studies on its functional properties. A full-length human laminin $\gamma 2$ chain cDNA containing 6 bp of the 5' UTR and 822 bp of the 3' UTR was constructed from four overlapping cDNA clones L52, HT2-7, L15 and L61 (Kallunki, P., Sainio, K., Eddy, R., Byers, M., Kallunki, T., Sariola, H., Beck, K., Hirvonen, H., Shows, T.B. & Tryggvason, K. (1992) J. Cell Biol. 119: 679-693). The resulting 4,402 bp cDNA was analyzed by restriction enzyme mapping and partial sequencing, and cloned into the pVL1393 recombinant transfer plasmid prior to transfer into the AcNPV- $\gamma 2$ baculovirus vector kindly provided by Max Summers (Texas A&M University). This baculovirus vector containing the human laminin $\gamma 2$ chain cDNA under the transcriptional control of the polyhedrin promoter was produced and purified following standard procedures, except that it was first enriched according to the method of Pen, et al. (Pen, J., Welling, G.W. & Welling-Wester, S. (1989), Nucl. Acid. Res. 17: 451) from the virus containing medium obtained by co-transfecting Sf9 cells with the wild-type virus (AcNPV) DNA and the recombinant transfer vector pVL 1393- $\gamma 2$. For expression of the recombinant protein, High Five (H5) cells were infected with the recombinant virus at a multiplicity of infection (MOI) of 5-10 pfu per cell by using standard protocols.

The recombinant $\gamma 2$ chain was purified by first resuspending the cells in 10 volumes of 50 mM Tris-HCl, pH 7.4, 100 mM NaCl, 2.5 mM EDTA, 1% Triton X-100, 1 mM PMSF and 1 mM NEM followed by homogenization in a Dounce homogenizer. The protein was extracted for 60 minutes on ice and solubilized proteins were removed by centrifugation at 1500 x g for 10 minutes at 4° C. The pellet was extracted again with buffer containing 1-3 M urea. The recombinant $\gamma 2$ chain was extracted with a buffer containing 5 M urea, and renatured by dialysis against 50 mM Tris-HCl, pH 7.4, 100 mM NaCl.

Preparation of Antibodies

Polyclonal antiserum against domain III of the laminin $\gamma 2$ chain was prepared and characterized as described previously. Briefly, rabbits were immunized s.c. four times using a $\gamma 2$ -GST fusion protein as antigen. The antigen contained 177 amino acid residues (res. # 391-567) from domain III of the $\gamma 2$ (SEQ ID NO:8) (Kallunki, P., Sainio, K., Eddy, R., Byers, M., Kallunki, T., Sariola, H., Beck, K., Hirvonen, H., Shows, T.B. & Tryggvason, K. (1992) J. Cell Biol. 119: 679-693). Antibodies against the GST-epitopes were removed from the antisera by negative immunoabsorption with GST-Sepharose made by coupling *E. coli* expressed GST protein to CNBr-activated Sepharose. The removal of anti-GST IgG was ensured by Western blotting analysis with GST-specific antibodies. The specificity of the antibody against the laminin $\gamma 2$ chain was also tested by Western blotting as well as by ELISA.

Polyclonal antibody against the C-terminus of the laminin $\gamma 2$ chain was produced in rabbits essentially as above for domain III using a $\gamma 2$ -GST fusion protein as antigen. The antigen contained 161 amino acids (res. # 1017-1178) from domain I/II of the $\gamma 2$ chain and antibodies against the GST-epitopes were removed from the antisera by negative immunoabsorption with GST-Sepharose. The specificity of the antibody was tested by Western blotting and ELISA.

Polyclonal antiserum against laminin-1 was a kind gift of Dr. Foidart (University of Liege, Belgium). Normal rabbit serum was obtained prior to immunization from the rabbits used for immunization. IgG from the laminin-1 and laminin $\gamma 2$ chain antisera, as well as from normal rabbit serum, was purified using Protein A Sepharose (Pharmacia, Uppsala, Sweden).

Cell Adhesion Assay

Microtiter plates (96 wells: Nunc, Copenhagen, Denmark) were coated with 100 μ l/well of laminin-1 (10 μ g/ml), laminin-5 (10 μ g/ml), or recombinant laminin $\gamma 2$ chain (10 μ g/ml) in PBS or 50 mM Tris-HCl, pH 7.4 by incubating the plates overnight at 4° C. Control wells were uncoated or coated with the same amounts of BSA. In some experiment the proteins were first denatured by dialysis overnight against 5 M urea, 50 mM Tris-HCl, pH 7.5 and then renatured by dialysis against 50 mM Tris-HCl, pH 7.5. Potential remaining active sites on the plates were blocked with 150 μ l of 10 mg/ml BSA in PBS for 2 hours at room temperature. The wells were

washed with PBS, and 100 ml of Eagle's MEM containing 5 mg/ml BSA was added. For the adhesion assays, KLN205 cells were detached from subconfluent cell culture dishes with trypsin-EDTA (0.25%-0.03%) and resuspended in Eagle's MEM/BSA (5 mg/ml) at a concentration of 2×10^5 cells/ml and allowed to recover for 20 minutes at 37° C. A total of 20,000 cells were then added to each well and allowed to attach for an additional 90 minutes at 37° C. The extent of cell adhesion was determined by measuring color yields at 600 nm, following fixation with 3% paraformaldehyde and staining with 0.1% crystal violet. For inhibition assays with the anti- $\gamma 2$ antibody, the substrate coated wells were incubated with 20 μ g/ml of anti- $\gamma 2$ chain IgG in PBS for 60 minutes prior to incubations with the cells.

Migration Assay

The effect of endogenous laminin-5 on migration of KLN205 cells was determined by using a modified Boyden chamber assay, as described by Hujanen and Terranova (Hujanen, E. & Terranova, V.P. (1985) Cancer Res. 45: 3517-3521), and the effect of exogenous laminin-5 by using a modified Transwell assay, as described by Pelletier, et al. (Pelletier, A.J., Kunicki, T. and Quaranta, V. (1996), J. Biol. Chem. 271:364).

The Boyden chamber assay was carried out as follows. Polycarbonate filters (pore size 10 μ m, diameter 12 mm; Costar, Cambridge, MA) were coated with 2.5 μ g of EHS type IV collagen, and used to separate the upper and lower compartments of the 50 μ l chamber. A total of 1×10^5 cells in Eagle's MEM containing 0.1% BSA were placed in the upper compartment, and the lower compartment was filled with medium with or without chemoattractants (50 μ g/ml laminin-1 or fibronectin). To study the effect of the laminin $\gamma 2$ chain antibodies on cell migration, anti- $\gamma 2$ (III) IgG or anti- $\gamma 2$ (C-term) IgG was added to the upper compartment together with the cells at a concentration of 20 μ g/ml. Normal rabbit IgG was used as a negative control. After an 8-hour incubation at 37° C in a humidified atmosphere, the filters were removed, fixed and stained (Diff-Quick, Baxter Diagnostics, Tübingen, Germany). The cells that had not migrated were removed from the upper surface of the filter with cotton swabs. Migration of cells was quantified by counting the cells on the lower surface of each filter in 10 randomly selected high power fields (x400). All assays were performed in triplicate.

The "Transwell" plate assay (Transwell plates with pore size 12 μ m, diameter 12 mm; Costar, Cambridge, MA) was used to determine the effect of exogenous laminin-5 on cell migration. The lower side of the membrane was coated with 2.5 μ g of EHS type IV collagen for 3 hours at room temperature. Both sides were blocked with 1% bovine serum albumin for 1
5 hour. A total of 1×10^5 cells were added per well in the upper compartment in Eagle's MEM containing 10% FCS, and the lower compartment was filled with 2.5 μ g/ml laminin-5 as a chemoattractant. Antibodies against the C-terminus and domain III of the γ 2 chains or nonimmune IgG were added to the upper compartment, together with the cells at a concentration 20 μ g/ml. Following a 16-hour incubation at 37° C the cells were fixed and stained. Cells on the
10 top surface of the membrane were removed with cotton swabs, and cells that had migrated to the lower side of the membrane were counted (12 fields \pm S.D.).

Immunohistochemical Staining

Five μ m thick paraffin sections were stained with polyclonal antibodies against laminin-1
15 or the γ 2 chain of laminin-5. In brief, the paraffin sections were first incubated with 0.4% pepsin in 0.1 M HCl at 37° C for 20 minutes to expose the antigens, blocked for nonspecific binding with 5% newborn rabbit serum, 0.1% BSA, and then incubated for 1 hour at 37° C with the polyclonal IgG diluted in TBS to 5-10 μ g/ml. Subsequently, a biotinylated swine-anti-rabbit antibody was applied, followed by incubation with a 1:400 dilution of Horseradish-Peroxidase-
20 Avidin-Biotin-Complex (DAKO, Copenhagen, Denmark). The color was developed in diaminobentsamidine (DAB), followed by counterstaining of the slides with hematoxylin.

Results

Characterization of Proteins and Epithelium-Derived Cells

25 Immunopurified trimeric laminin-5, isolated from the culture medium of HaCat cells contained two major bands when analyzed by SDS-PAGE. These bands corresponded, respectively, to the 165 kDa γ 2 chain, and the 155 kDa and 140 kDa γ 2 and β 3 chains migrating as a single band, as reported previously. Additionally, a weak band of about 105 kDa corresponding to the processed γ 2 chain could be observed.

Full-length human recombinant laminin γ 2 chain was produced in High-5 *Spodoptera frugiperda* insect cells using the baculovirus system. Since the γ 2 chain was not secreted to the culture medium, possibly because it was not assembled intracellularly into a normal heterotrimer, it was isolated from the cell fraction as described in *Materials and Methods*. The protein was
5 extracted under denaturing conditions using 5 M urea, renatured by extensive dialysis against 50 mM Tris-HCl, 100 mM NaCl, pH 7.4, and purified. The purified recombinant γ 2 chain was full length (approximately 155 kDa) and highly pure as determined by SDS-PAGE analysis.

The HaCat human keratinocytes and mouse KLN205 squamous carcinoma cells were shown to express laminin-5, based on Northern blot analyses and immunostaining, using a
10 cDNA probe and/or polyclonal antibodies specific for the γ 2 chain, respectively. Furthermore, the KLN205 cells developed γ 2 chain positive primary tumors and metastases in mice *in vivo* (data not shown). Following intramuscular or subcutaneous inoculations, large primary tumors developed in 4 weeks with numerous lung metastases after 4-6 weeks. KLN205 cells injected into the tail vein produced multiple lung tumors (experimental metastases) in four weeks.
15 Consequently, both cell types were considered appropriate for the cell attachment and migration experiments carried out in this study.

Laminin-5 Molecule, but not Recombinant Laminin γ 2 Chain, Promotes Cell Adhesion

The laminin-5 and recombinant γ 2 chain prepared in this study, as well as commercial
20 laminin-1, were used as substrata in attachment assays (**FIGURE 1**) with the two epithelium-derived HaCat and KLN205 cell lines that both express laminin-5. Both cell lines attached about 2.5 times more readily to laminin-1 than to plastic. Adhesion of the cells to laminin-5 appeared to be slightly higher than that to laminin-1, but the differences were not statistically significant. The cells attached equally well to laminin-5 preparations denatured in 5 M urea and then
25 renatured by dialysis against 50 mM Tris-HCl, 100 mM NaCl, pH 7.4, as described for the recombinant γ 2 chain above, indicating that this treatment did not affect the binding properties of the trimeric molecule. The attachment to laminin-5 did not significantly decrease in the presence of two different polyclonal antibodies made against the short or long arms of the γ 2 chain or pre-IgG. Different amounts of the antibody against the short arm of the γ 2 chain were also tested (up
30 to 50 μ g/ml), but no effects on cell adhesion were observed. When the cells were plated on the

recombinant $\gamma 2$ chain alone, the attachment was not significantly higher than that to plastic, this attachment not being influenced by polyclonal antibodies against the $\gamma 2$ chain. The data confirm previous results showing that trimeric laminin-5 promotes adhesion of epithelial cells, but the present results further strongly suggest that this adhesion is not mediated by the $\gamma 2$ chain.

Antibodies Against Laminin $\gamma 2$ Domain III, But Not Domain I/II, Inhibit Cell Migration

The potential role of the $\gamma 2$ chain of laminin-5 in cell migration was examined for the KLN205 cells *in vitro* using Boyden and Transwell chamber assays as described in *Materials and Methods*.

Migration was first studied in the Boyden chamber assay using laminin-1 and fibronectin in the lower chamber as chemoattractants (See **Figure 2A**). The two compartments of the chemotactic Boyden chambers were separated by a type IV collagen coated porous filter (pore size 8 μm). The cells (1×10^5) in MEM containing 0.1% BSA were placed in the upper compartment, and laminin-1 (+/-) or fibronectin (-/+) in MEM containing 0.1% BSA were added as chemoattractants to the lower compartment. IgG against $\gamma 2$ chain domains III, I/II or preimmune IgG was added to the upper compartment with the cells at a concentration of 20 $\mu\text{g/ml}$. After an 8-hour incubation at 37°C the filters were removed and migration of cells to the lower surface of the filter was quantitated. The data are expressed as percentage of migrated cells (+/- SD (bars)) per high power field, setting migration in the presence of pre-immune IgG as 100%. Cells were counted in ten randomly selected high power fields to triplicate assays. When polyclonal IgG against the short arm of the $\gamma 2$ chain was added to the upper compartment containing the cells, the migration of cells through the filter was decreased to about 35 to 45% of that observed with the preimmune serum. In contrast, the polyclonal IgG against C-terminal domain I/II did not affect migration of the cells.

The effects of the two antibodies were similarly used in the Transwell assay using native laminin-5 as chemoattractant in the lower compartment (See **Figure 2B**). The lower side of the membrane was coated with EHS type IV collagen, and the lower compartment was filled with 2.5 $\mu\text{g/ml}$ laminin-5 as a chemoattractant. Pre-immune IgG, IgG against the $\gamma 2$ chain domains III or I/II were added to the upper chamber containing the cells. Following a 16-hour incubation the cells were fixed and cells at the lower side of the membrane were counted (12 fields +/- SD).

The results were essentially the same as in the Boyden chamber assay. Thus, addition of IgG raised against domain III of the $\gamma 2$ chain inhibited the migration to about 50% as compared with preimmune IgG, while the polyclonal IgG against domain I/II did not affect the cell migration.

These *in vitro* results demonstrate that laminin-5 have a role in the locomotion of epithelium-derived cells, and that this function can be inhibited by antibodies directed against domain III of the $\gamma 2$ chain.

Thus, antibodies against the short arm of the laminin $\lambda 2$ chain inhibited the migration of KLN205 squamous carcinoma cells by about 55-65% as determined in the Boyden chamber migration assay. Interestingly, the antibodies used here were directed against 177 amino acid residues of domain III (SEQ ID NO:8) that when deleted by mutation cause lethal junctional epidermolysis bullosa. Accordingly, the short arm of the laminin $\lambda 2$ chain is important for the interaction of this laminin isoform to other extracellular matrix proteins and this interaction is also involved in the cell migration process.

EXAMPLE 2

The following example describes, in detail, the preparation of monoclonal antibodies according to the invention as well as demonstrating their use in inhibiting tumor cell growth in laminin-5 secreting tumors.

Monoclonal antibodies against the $\gamma 2$ chain of laminin-5 were produced by immunizing Balb/c mice with 100 ug GST-laminin- $\gamma 2$ -III fusion protein as antigen. The GST-laminin- $\gamma 2$ -III fusion protein contains human laminin- $\gamma 2$ -chain amino acid residues 391-567 (SEQ ID NO:8). Subsequent to immunization, spleen cells from the immunized mice were fused with mouse myeloma cell obtained from cell line P3X63Ag.8.653 (ATCC #CRL-1580). The hybridoma clones were then screened in immunohistology on frozen and paraffin sections (human cervix carcinoma, normal cervix and normal skin) for the production of the anti-laminin- $\gamma 2$ antibody. The staining result was compared to negative control, mouse normal serum and IgG, and to the positive result obtained with well-characterized anti-laminin-5, $\gamma 2$ chain polyclonal antibody (described in Pyke, et al., 1995). The hybridoma clones were also screened in ELISA. The best hybridoma clones were picked and cloned again twice (single cell cloning) to ensure that the produced hybridoma cell line was monoclonal.

The following describes the details of the production of three specific hybridoma clones

and corresponding monoclonal antibodies produced therefrom. Characterization studies were conducted with respect to the 4G1, 5D5 and 6C12 monoclonal antibodies. Western blot analysis and ELISAs were carried out to confirm the specificity of the antibodies to the $\gamma 2$ chain of laminin 5. Western blot analysis involved running recombinant laminin 5 $\gamma 2$ chain (as well as appropriate controls) in an SDS-PAGE gel, blotting the gel on a nylon membrane, and incubating the membrane with the antibodies

For ELISA, plates were coated with 100 μ l GST- $\gamma 2$ -III fusion protein (antigen) (Salo et al., Matrix Biology 18:197-210 (1999) at a concentration of 2.5 μ g/ml in 0.1M carbonate/bicarbonate buffer (pH 9) overnight at 4° C (0.25 μ g/well). The ELISA plate was then washed three times with a PBST solution (200 μ l) (10mM potassium phosphate, 150 mM NaCl), pH 7.5, and 0.05% Tween-20. Non-specific binding was then blocked by addition of BSA-PBS (1% bovine serum in PBS buffer (10mM K₃P0₄, 150 mM NaCl, pH 7.5)) (200 m/well) for a period of 90 minutes. To this, a dilution of negative controls (normal mouse serum) and a sample diluted in BSA-PBS (Mab 4G1, 5D5 or 6C12) were added and then the ELISA plate was incubated for 1 hour at room temperature. After incubation, the ELISA plate was then washed with PBST three times. Next, HRP-conjugated anti-mouse IgG secondary antibody (Peroxidase (HRP) conjugated Rabbit Anti-Mouse IgG (H+L), Jackson Laboratories #315-035-045) was added and the plate was incubated at room temperature for 30 minutes. The ELISA plate was then washed again three times with PBST solution (200 ml). An ABTS-peroxide substrate was then added to the wells (ABTS diluted in 0.1 M Na-citrate, pH5; diluted immediately before assay use 1ml to 10ml with Na citrate buffer + 2 μ l 30% hydrogen peroxide) and then the plate was allowed to incubate in the dark for 30 minutes. The absorbance was then read with a micro plate reader at 405 nm at 30 and 60 minutes.

These analyses demonstrated the specificity of the monoclonal antibodies for domain III of the laminin 5 $\gamma 2$ chain. Epitope mapping of the epitopes recognized by Mab 4G1, 5D5 or 6C12 indicated that they each bound epitopes within the amino acid sequence of SEQ ID NO:6 (which is a portion of domain III of the $\gamma 2$ chain that lacks part of the amino and carboxy terminal portions of domain III), and did not bind to epitopes within the amino acid sequence of SEQ ID NOS: 9 and 10 (the 9 amino terminal and 41 carboxy terminal amino acids of domain III, respectively).

Monoclonal antibodies against the $\gamma 2$ chain of laminin-5 were then tested for efficacy in inhibiting tumor cell growth in laminin-5 secreting tumors.

Study 1: Tumor Growth in Immunosuppressed Mice

The following study demonstrates the ability of IgG immunoglobulin against human laminin-5, $\gamma 2$ -III-domain (Mab 5D5) to affect the number and size of metastases in immune deficient mice.

10^6 human squamous epithelial carcinoma cells were injected into the tail vein of immunosuppressed mice for tumor implantation. The cell lines used were human squamous epithelial carcinoma cells, cell line A431 and HSC-3. The cells were provided in suspensions in a medium containing DMEM-glutamax, 1% penicillin-streptomycin, 1% Na-pyruvate, 5% FCS. The cells were re-suspended in sterile Ca and Mg free PBS for inoculation. A control cell count was performed for the cell suspension at arrival and the cell density and the injected volume was recorded. The origin of the cells is HSC-3: Japan Health Science Research Resources Bank, JCRB 0623 A431: ATCC catalog number CRL-1555. The immunosuppressed mice were selected as they are susceptible to grow cells of human origin as is well known in the field. The tumor cells in groups 3 and 6 were injected into mice with test item (test item was 50 μ g/ml) for tumor implantation. The tumor cells were allowed to grow for one week after which the animal received intravenous injections of the test item twice a week for four weeks.

Table 1. Study Layout

Group	Mouse Strain		Animal Number	Cell Line	Treatment
1	Balb/c~nudet	5	1-5	-	-control, no treatment
2	Balb/c-nude ¹	5	6-10	HSC-3	+control, no treatment
3	Balb/c-nude ¹	5	11-15	HSC-3	Test item treatment: 50 μ g 5D5/mouse injection
4	SCID ²	5	21-25	-	-control, no treatment
5	SCID~	5	26-30	A431	+control, no test item
6	SCIDZ	5	31-35	A431	Test item treatment: 50 μ g 5D5/mouse injection

¹ Balb/c-nude (BALBicABom-nu, M&B A/S, Denmark)

² Fox Chase Scid (C.B-17/lcr scid/scid, M&B A/S, Denmark) immunodeficient mice.

After the treatment period, the animals were killed and tissue samples were collected. Number and size of the tumors in different tissues were counted and compared.

5 *Test Items and Dosing Solutions*

The test item was IgG immunoglobulin against human laminin-5, $\gamma 2$ -III-domain (Mab 5D5). The test item was produced with monoclonal hybridoma method *in vitro* as set forth above. The test item (Mab 5D5) was suspended in sterile phosphate buffered saline (PBS) with a concentration of 1 mg/ml. The vehicle was sterilized using a 0.2 μ m filter. The delivered test
10 item was diluted with sterile PBS 50:50 to give a dosing concentration of 500 μ g/ml.

The test item was administered intravenously into the lateral tail vein of the immunosuppressed mice in a volume of 0.1 ml/animal. The dosing was twice a week on Mondays and Thursdays. The first dose of test item was administered one week after the induction of experimental metastasis.

15 After four weeks of treatment (eight doses of test item), the animals were killed by exsanguination with cardiac puncture in CO₂ anesthesia. Blood was collected and serum separated and frozen in -20° C. A gross necropsy was performed and the macroscopic signs were recorded with special attention to macroscopic tumor masses, which were calculated and measured if possible. The following organs/tissues were collected and weighed: lungs, lymph
20 nodes (cervical and mesenteric), liver, and spleen. The organs/tissues were rinsed in PBS and fixed in 4% phosphate buffered formalin.

Clinical Signs

Animal number 6 had a thickening of the tail from day 5 through the whole study. The
25 tail of animal number 11 turned dark/black after tumor cell inoculation and eventually turned necrotic. Half of the tail was missing from day 7 onward. No other treatment related clinical signs were recorded. One animal (number 8, group 2) was found dead on the morning of the day following tumor cell inoculation. Gross necropsy did not reveal any macroscopic changes. All
30 other animals survived in good condition during the whole study.

Necropsy

The injected tumor cells induced tumor growth almost only in the lungs. Other tissues with macroscopic metastases include spleen, liver, small intestine, and preputial gland. The SCID mice had changes in the liver which might be of microbial origin. In the lungs, the metastases were so numerous and so small that it was impractical to calculate or measure individual metastases.

The following Table 2 represents a summarization of the results of the mice treated from Table 1.

Table 2. Experimental Metastases in Lung

Group	Mouse Strain	N	Cell Line	Treatment	Number of Mice with Macroscopic Lung Metastases Observed
1	Balb/c-nude	5	-	-control, no treatment	-
2*	Balb/c-nude	5	HSC-3	+ control, no treatment	4/4 (full of mastastases)
3	Balb/c-nude	5	HSC-3	Test item treatment	1/5
4	SCID ~	5	-	-control, no treatment	-
5	SCID	5	A431	+control, no test item	3/5
6	SCID	5'	A431	Test item treatment	4/5

* one mouse was dead at the end of the second study

As can be seen from Table 2 above, the treated Balb/c-nude mice had 1 of 5 mice with macroscopic lung metastases while 4 of 4 untreated control Balb/c-nude mice had macroscopic lung metastases.

EXAMPLE 3

Monoclonal antibody 5D5 was tested against HT29 carcinomas in a tumor growth inhibition assay. The assay compared immunotherapy with 75 and 25 µg/mouse 5D5, qod x 15, to conventional chemotherapy with 100 mg/kg CPT-11 (irinotecan/Campostar), qwk x 3.

Methods and Materials

Female nude athymic mice (Harlan) were 13 weeks of age on day 1 of the study. The

animals were fed *ad libitum* water (reverse osmosis, 1 ppm Cl) and the NIH 31 Modified and Irradiated Lab Diet® consisting of 18.0% protein, 5.0% fat, and 5.0% fiber. Mice were housed in static microisolators on a 12-hour light cycle at 21-22 ° C (70-72 ° F) and 40%-60% humidity.

5 Tumor Implantation

An HT29 carcinoma fragment (1 mm³) was implanted subcutaneously in the flank region of each mouse. When the tumors reached a size ranging from 62.5-126 mg, the mice were sorted into five treatment groups to provide a group mean tumor weights of 84.2-85.5 mg. Estimated tumor weight was calculated using the formula:

$$\text{Tumor Weight (mg)} = \frac{w^2 \times l}{2}$$

Where *w* = width and *l* = length in mm of the HT29 carcinoma.

Dosing solutions of 5D5 and control IgG were prepared fresh daily by dilution with phosphate-buffered saline. CPT-11 (Pharmacia; 20 mg/mL) was diluted with saline on each day of dosing.

On day 1, mice were sorted into five groups of animals (*n* = 10/group), and dosing was initiated according to the protocols listed in Table 3.

Table 3. Protocol Design for the HT29-e29 Study

Group	n	Treatment Regimen I			
		Agent	mg/kg	Route	Schedule
1	10	No treatment	n/a		
2	10	CPT-11	100	IP	QwKx3
3	10	Control IgG	75 ug/ mouse	IV	Qod x 15
4	10	5D5	75 ug/ mouse	IV	Qod x 15
5	10	5D5	25 ug/ mouse	IV	Qod x 15~

As a positive reference drug, CPT- 11 was administered once per week for three weeks (qwk x 3) in 100 mg/kg doses. CPT- 11 was delivered i.p. in volumes of 0.2 ml/20 g body weight, which were body-weight adjusted. Doses of 5D5 or control mouse IgG were delivered intravenously in volumes of 0.2 mL/mouse. The antibody doses were not body-weight adjusted.

5 Untreated Group I mice served as controls for the CPT-11 therapy. Group 3 mice received 15 µg/mouse doses of control IgG once daily on alternate days (qod x 15). Mice in groups 4 and 5 received 75 and 25 ug/mouse doses of 5D5 x 15, respectively.

Endpoint

10 Efficacy was evaluated in a tumor growth inhibition assay. Tumors were measured twice weekly until the study was terminated on day 31. Each animal was then euthanized and its HT29 carcinoma was excised and weighed. Treatment may produce complete tumor regression (CR) or partial tumor regression (PR) in an animal. In a CR response, there is no measurable tumor mass at the completion of the study. In a PR response, the tumor weight is lower than the weight

15 on day, but greater than 0 mg. All tumors that did not regress were included in the calculation of tumor growth inhibition.

The increase in tumor weight for each animal was calculated as the difference between the actual tumor weight at the end of the study and the calculated tumor weight on day 1. These values were used to calculate the group mean tumor weight increases. Tumor growth inhibition

20 was calculated from the group mean tumor weight increases of treated and control mice by the following equation:

$$\%TGI = \left[1 - \left(\frac{\text{MeanNetTum or Weight}_{\text{Treated}}}{\text{MeanNet Turn or Weight}_{\text{control}}} \right) \right] \times 100 \%$$

Toxicity

The mice were weighed twice weekly until the end of the study. They were examined frequently for clinical signs of any adverse, drug-related side effects. Acceptable toxicity for cancer drugs in mice is defined by the NCI as a mean group weight loss of less than 20% during

30 the test, and not more than one toxic death among ten treated animals.

Statistics and Graphical Analyses

The unpaired t-test and Mann-Whitney U-test (for analysis of means and medians, respectively) were used to determine the statistical significance of the difference in mean tumor weights for mice in a treatment group and mice in a control group. The two-tailed statistical analyses were conducted at $P = 0.05$.

Results

Efficacy: Growth of HT29 Colon Carcinomas in Control Mice

Treatment protocols are listed in Table 3. Group 1 mice received no treatment and served as controls for CPT-11 and 5D5 therapy. Group 3 mice received fifteen 75 µg/mouse doses of irrelevant mouse IgG on alternate days (qod x 15). Table 4 summarizes the results for all groups in the study. The mean values for actual day 31 tumor weights in untreated and IgG-treated mice are 640.0 and 696.2 mg, respectively.

Table 4. Treatment Response Summary for the HT29-e28 Study

Gp		Regimen I				Final Tumor Weight Mean \pm SEM (n)	Tumor Growth Inhibitio n	# CP	Mean % Tumor Decrease	# CR	Max. % BW Loss Day	# Death ^a	
		Agent	mg/kg	Route	Schedule							TR	NT R
1	10	No treatment	n/a			640.0 \pm 124.9 mg (10)	0%	0	None	0	-0.4%; Day 4	0	0
2	10	CPT-11	100	IP	Qwk x 3	447.9 \pm 91.8 mg(10)	34.7%	0	None	0	-5.8%; Day4	0	0
3	10	Control IgG	75 µg/ mouse	IV	Qod x 15	696.2 \pm 131.4 mg (10)	0%	0	None	0	-3.6%; Day 4	0	0
4	10	5D5	75µg/ mouse	IV	Qod x 15	543.5 \pm 149.3 mg (8)	17.8%		None	0	-2.0%; Day 4	0	0
5	10	5D5	25µg/ mouse	IV	Qod x 15	700.7 \pm 116.1 mg(10)	0%	0	None	0	-1.2%; Day4	0	0

Response of HT29 Xenografts to Intraperitoneal CPT-11 Therapy

Group 2 mice were treated once weekly for three weeks (qwk x 3) with i.p. injections of 100 mg/kg CPT-11 (Table 3). No tumors regressed in response to CPT-11. The final mean tumor weight in Group 3 mice was 447.9 mg (Table 4). Group 2 mice experienced 34.7% tumor growth inhibition, relative to the untreated mice. This result, which is illustrated in a bar graph in **FIGURE 3**, was not statistically significant ($P = 0.23.11$, unpaired two-tailed t-test). **FIGURES 4A-E** shows the growth of individual tumors in all treatment groups, as calculated from caliper measurements. CPT-11 treatment caused a decrease in the slope of tumor growth.

Response of HT29 Xenografts to Intravenous 5D5 Immunotherapy

5D5 was administered intravenously to mice in Groups 4 and 5 on the qod x 15 schedule at 75 and 25 ug/mouse, respectively (Table 3). No tumor regressions were observed. The 75 and 25 ug/mg mouse 5D5 treatments yielded final actual mean tumor weights of 543.5 and 700.7, respectively (Table 4). The high dose of 5D5 inhibited HT29 carcinoma growth by 17.8%, relative to tumor growth in untreated mice. Tumor growth inhibition in Group 4 mice, relative to untreated and IgG-treated mice, was not statistically significant ($P = 0.6241$ and 0.453 , respectively; t-test). Group 5 mice experienced no inhibition of tumor growth. **FIGURE 3** illustrates the lack of significant tumor growth inhibition, given the large error (SEM) bars. **FIGURES 4 A-E** shows that there was a modest decrease in the slopes of the tumor growth curves in animals treated with 75 ug/mouse 5D5.

Side Effects

All therapies were well tolerated. The highest group mean body-weight loss, an acceptable 5.8%, was recorded in mice treated with CPT-11. Body weight losses in antibody-treated mice were 3.6% or lower.

Discussion

The HT29 colon carcinoma xenograft model was appropriate for 5D5 evaluation because HT29 cells produce laminin. Growth of primary tumors can be impeded by anti-proliferative agents, such as CPT-11, as well as by agents that prevent invasion of the substratum. Combinational

treatments using monoclonal antibodies against the $\gamma 2$ chain of laminin-5, such as 5D5, with anti-proliferative agents such as CPT-11 are also contemplated as part of the invention. Treatment efficacy was based on tumor growth inhibition, i.e., the difference between the mean increase in tumor size in control and treated groups of animals during the 31-day study. Although there was no response to 5D5 at a dose of 25 $\mu\text{g}/\text{mouse}$, tumor growth was inhibited by 17.8% at 75 $\mu\text{g}/\text{mouse}$ (Table 4 and **FIGURE 3**). Thus, a 75 $\mu\text{g}/\text{mouse}$ dose of 5D5 produced some therapeutic effect against HT29 colon carcinomas. In general, there was a reduction in the slopes of the tumor growth curves in mice treated with CPT-11 and 5D5 (**FIGURES 4A-E**). Accordingly, these results indicate that anti-laminin immunotherapy has application in cancer treatment of laminin-5 secreting tumors.

in summary, established HT29 colon carcinomas responded to therapy with 75 $\mu\text{g}/\text{mouse}$ doses of 5D5. High dose 5D5 immunotherapy achieved 50% of the tumor growth inhibition that was produced by CPT-11 chemotherapy. The tumor growth shown in **FIGURES 4A-E** curves suggest that 5D5 immunotherapy can impair colon tumor growth at doses of 75 $\mu\text{g}/\text{mouse}$ or higher.

Those skilled in the art will know, or be able to ascertain, using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. These and all other equivalents are intended to be encompassed by the following claims.

We claim:

1. Use of an antibody that binds to one or more epitopes of a laminin 5 γ 2 chain for the preparation of a medicament for inhibiting tumor growth in a patient with a tumor.
2. The use of claim 1 wherein the antibody is a monoclonal antibody.
- 5 3. The use of claim 1 or 2 wherein the tumor is a carcinoma.
4. The use of any one of claims 1-3 wherein the medicament further comprises one or more further anti-tumor agent.
5. The use of claim 4 wherein the further anti-tumor agent is a chemotherapeutic agent.
- 10 6. An isolated antibody that binds to one or one or more epitopes of domain III of the laminin 5 γ 2 chain but does not bind to epitopes within the amino acid sequence of **SEQ ID NOS: 9 and 10.**
7. The isolated antibody of claim 5 wherein the antibody is a monoclonal antibody.
8. A pharmaceutical composition comprising the isolated antibody of claim 5 and a pharmaceutically acceptable carrier.
- 15 9. Isolated hybridoma cells that express the monoclonal antibody of claim 6.
10. Use of the isolated antibody of any one of claims 6-8 for the preparation of a medicament for one or more of inhibiting tumor growth, inhibiting tumor metastasis, and detecting invasive cells in a patient with a tumor.
- 20 11. A pharmaceutical composition comprising an antibody that binds to one or more epitopes in domain III of laminin 5 γ 2 chain and a further anti-tumor agent.
12. The pharmaceutical composition of claim 6 wherein the further anti-tumor agent is a chemotherapeutic agent.
- 25 13. A method for detecting the presence of invasive cells in a tissue comprising contacting the tissue sample with the antibody of any one of claims 6-8 to form an immunocomplex, and detecting formation of the immunocomplex, wherein the formation of the immunocomplex correlates with the presence of invasive cells in the tissue.

30

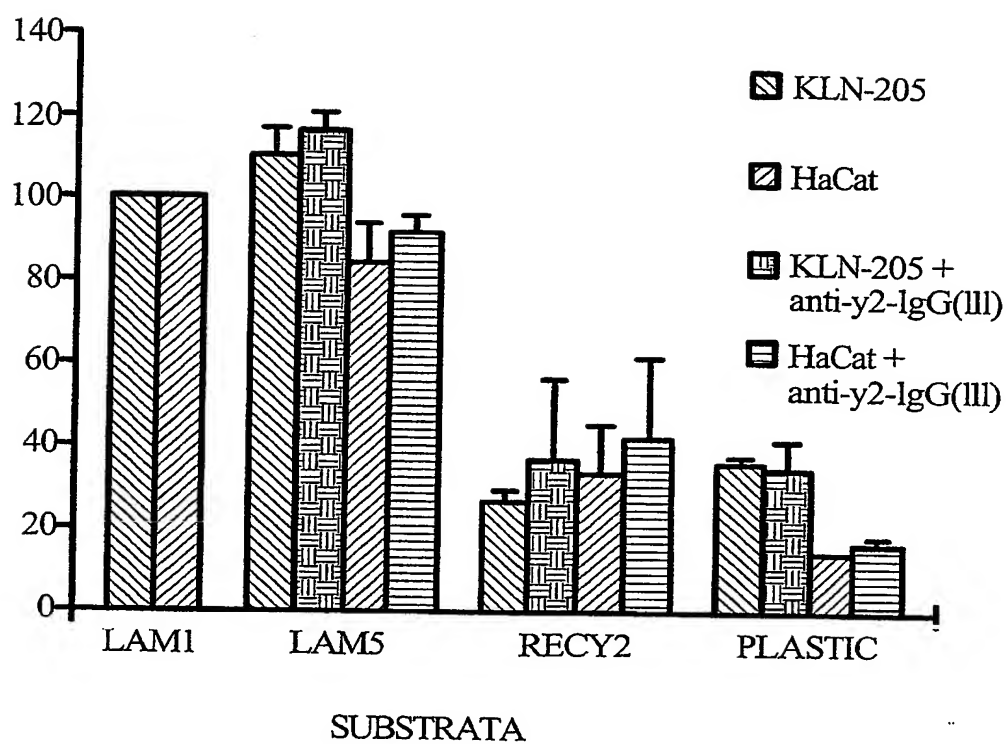
FIG. 1

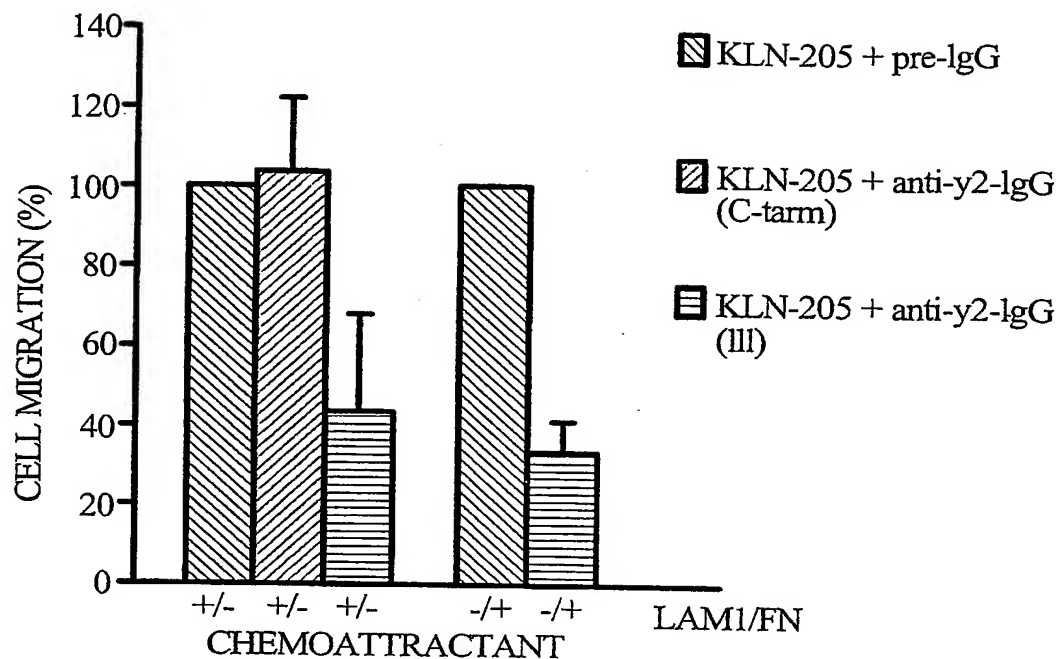
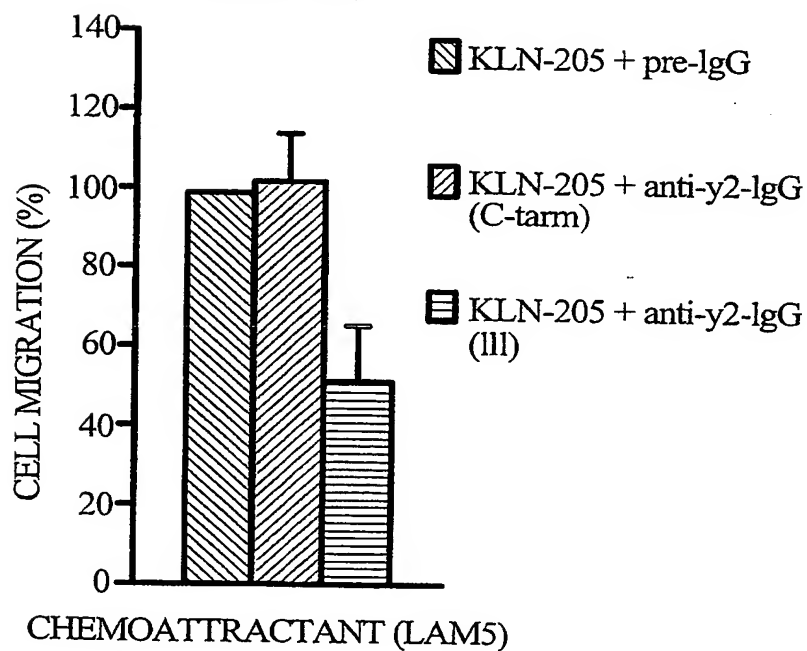
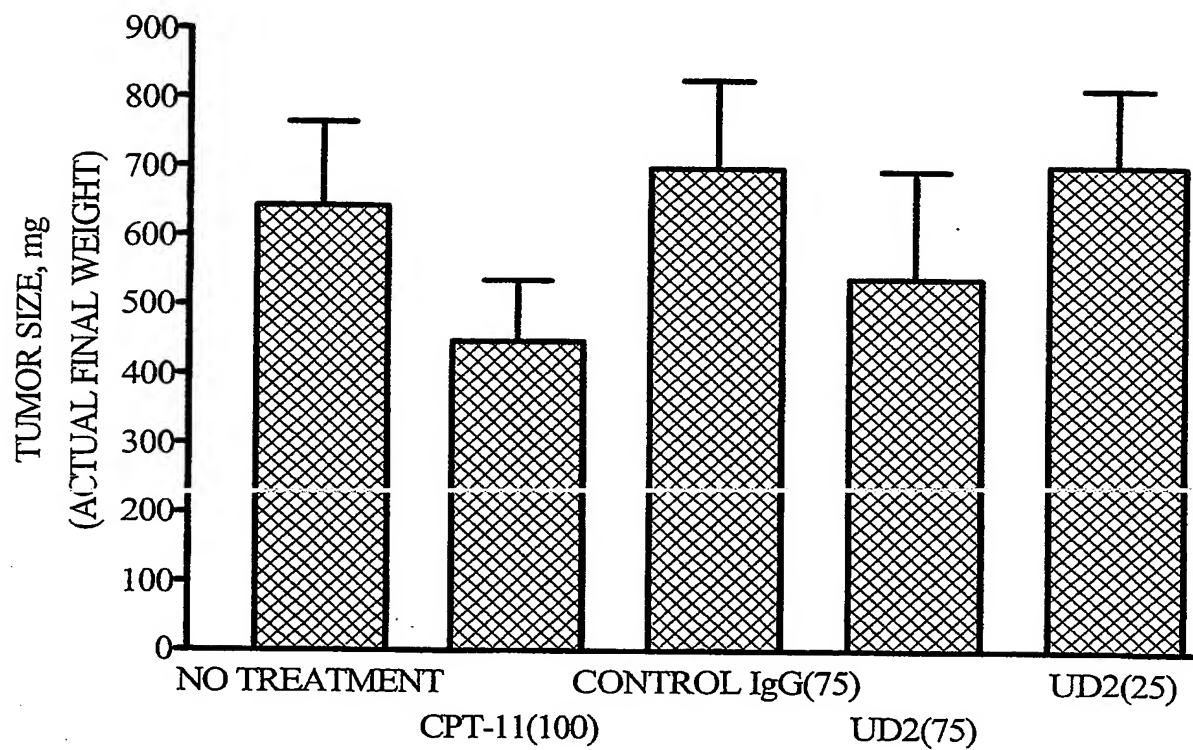
FIG. 2A*FIG. 2B*

FIG. 3

TUMOR GROWTH INHIBITION ON DAY 31
IN THE HT29-e28 STUDY



Tumor Growth Curves for Individual Mice in the HT29-e28 Study Group 1: No Treatment

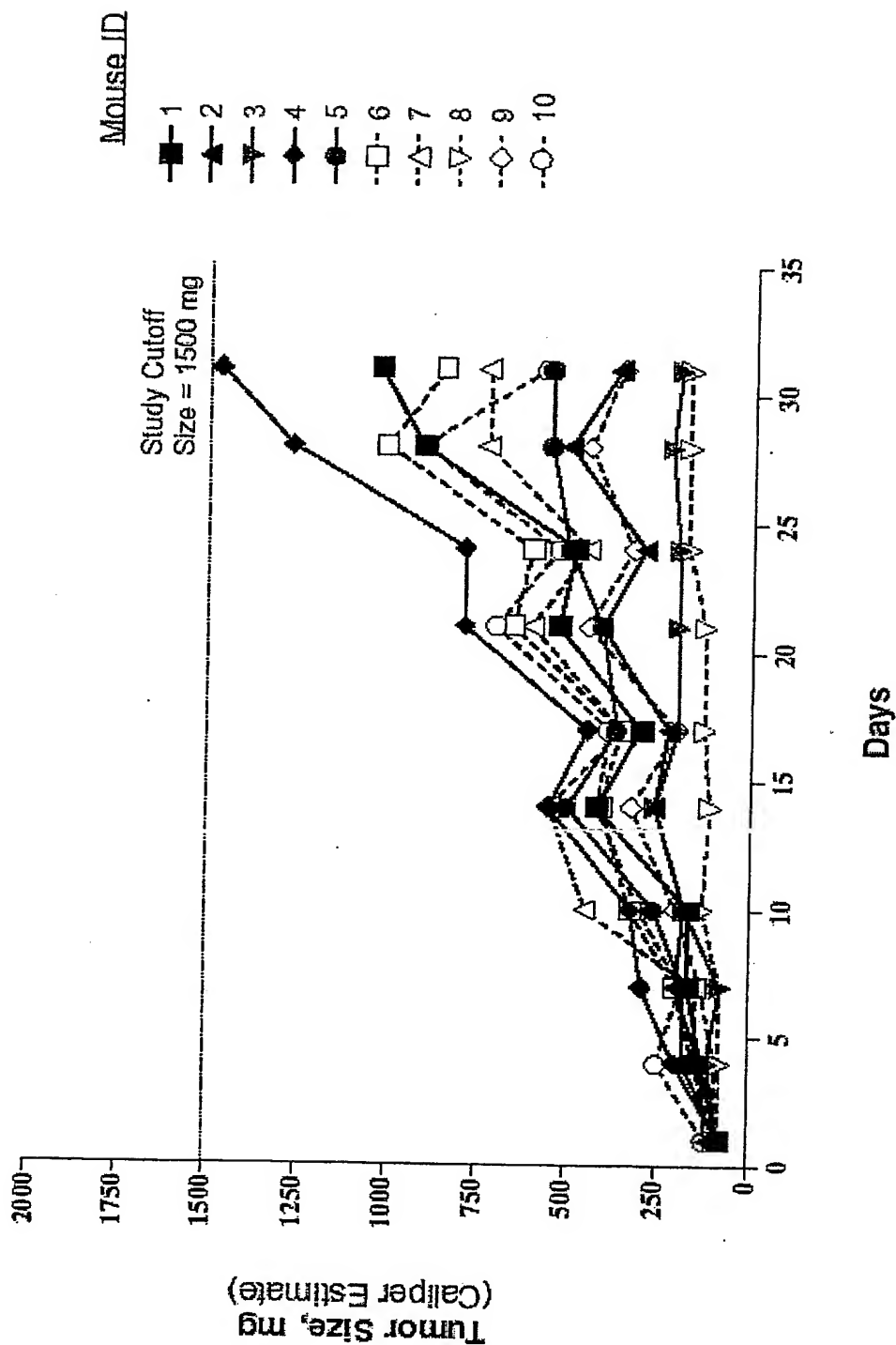


FIGURE 4A

Tumor Growth Curves for Individual Mice in the HT29-e28 Study
Group 2: CPT-11: 100 mg/kg; Ip: qwk x 3

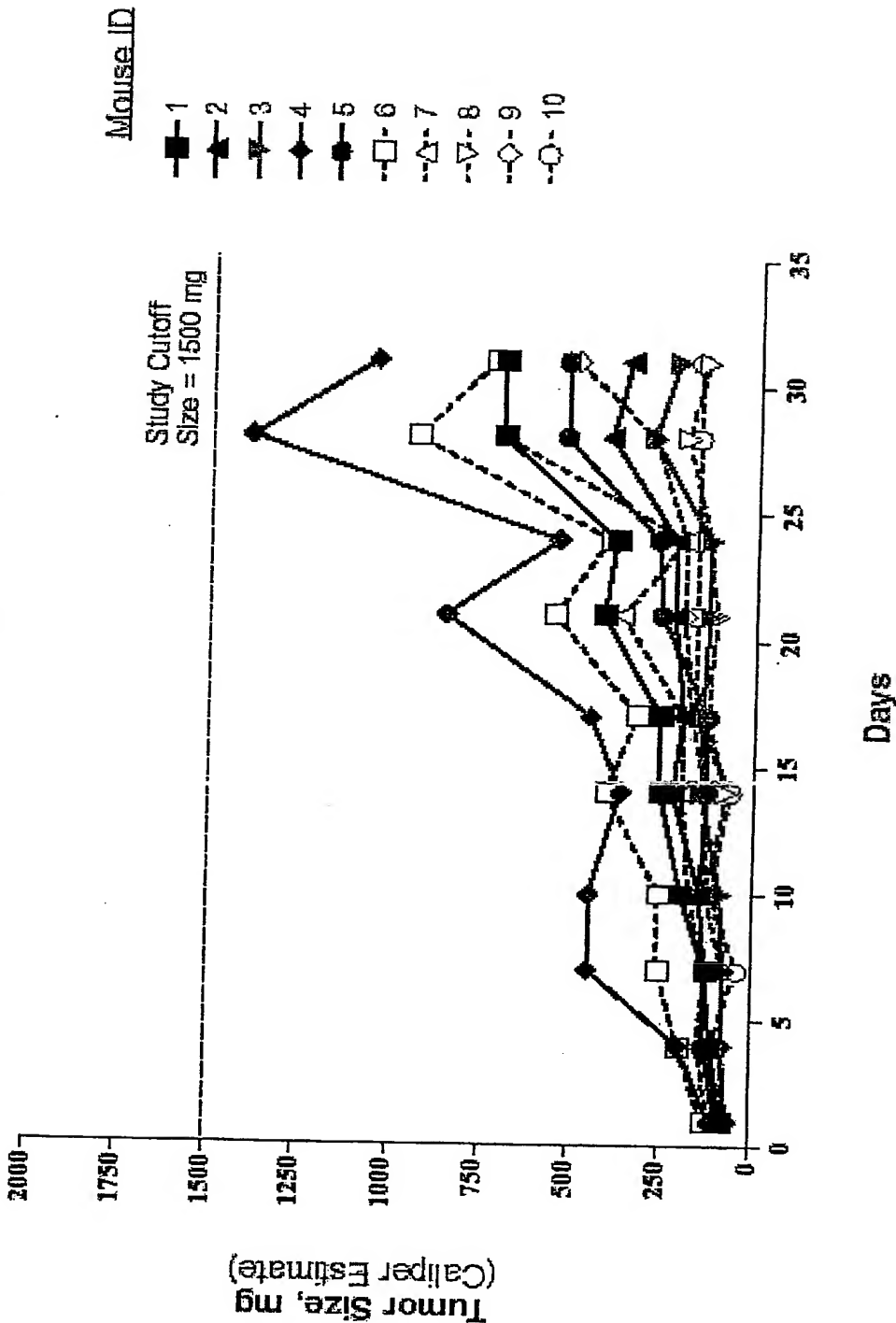


FIGURE 4B

Tumor Growth Curves for Individual Mice in the HT29-e28 Study
Group 3: Control IgG; 75 mg/kg; iv; qod x 15

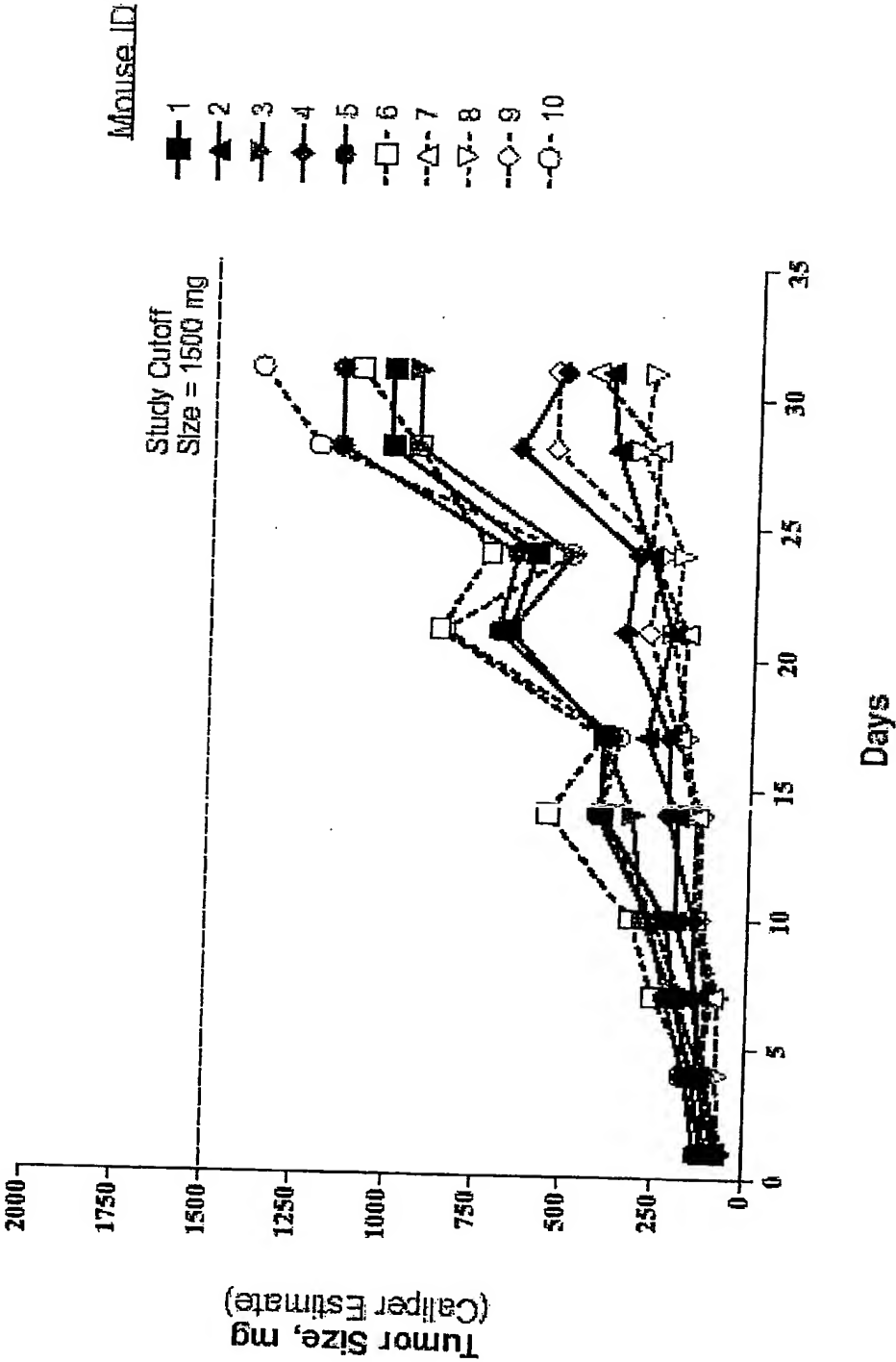


FIGURE 4C

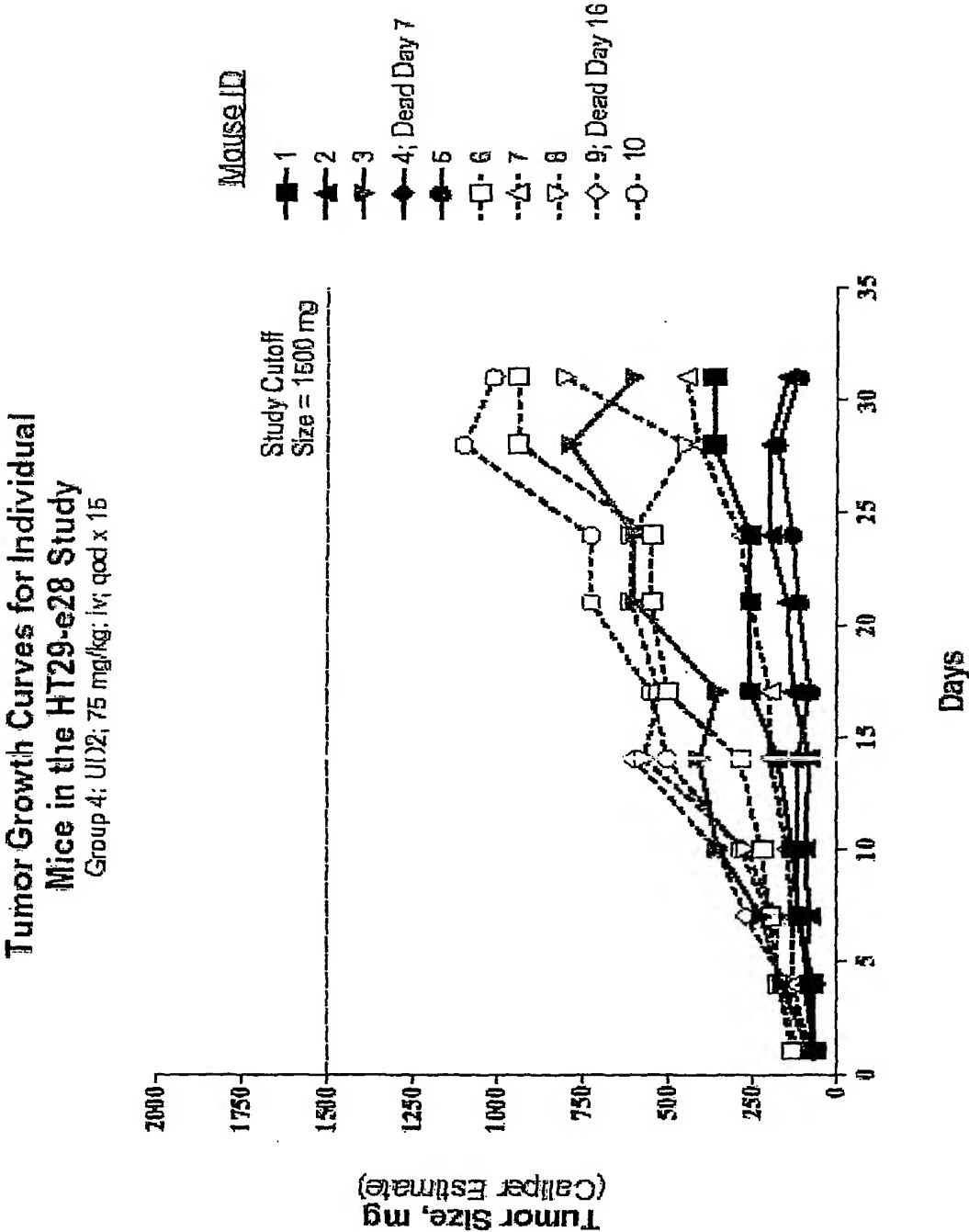


FIGURE 4D

Tumor Growth Curves for Individual Mice in the HT29-e28 Study
Group 5: UD2; 25 mg/kg; iv; qod x 15

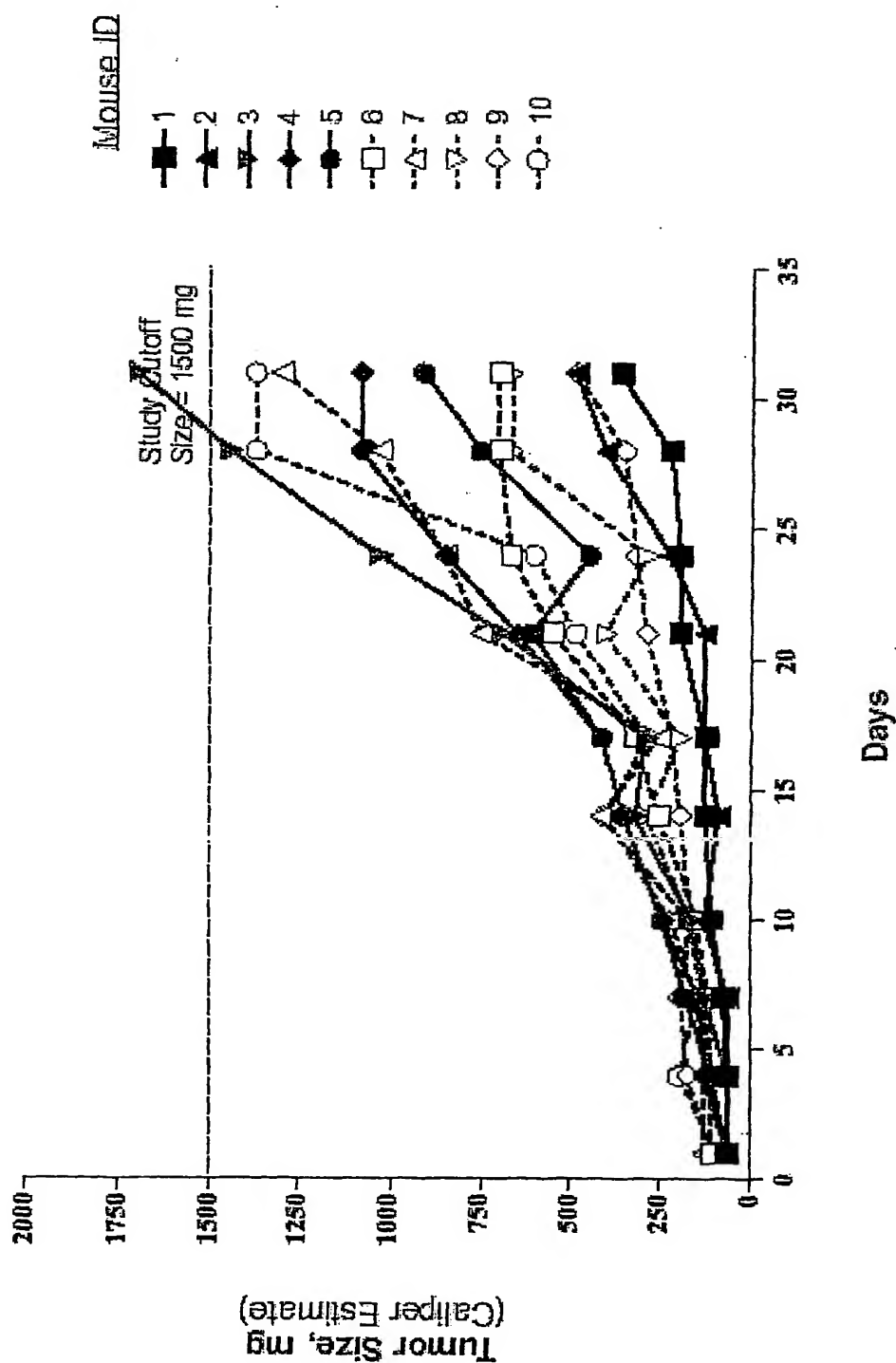


FIGURE 4E

SEQUENCE LISTING

<110> Tryggvason, Karl
Salo, Sirpa

<120> Use of antibodies to the gamma 2 chain of laminin 5 to inhibit tumor growth and metastasis

<130> 02-1147-PCT

<150> 60/422,009

<151> 2002-10-29

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Met Pro Ala Leu Trp Leu Gly Cys Cys Leu Cys Phe Ser Leu Leu Leu

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Lys Ser Arg Gln Cys Ile Phe Asp Arg Glu Leu His Arg Gln Thr Gly

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Asn Gly Phe Arg Cys Leu Asn Cys Asn Asp Asn Thr Asp Gly Ile His

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Cys Glu Lys Cys Lys Asn Gly Phe Tyr Arg His Arg Glu Arg Asp Arg

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Asp Asn Ser Gly Arg Cys Ser Cys Lys Pro Gly Val Thr Gly Ala Arg

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Leu	Glu	Gly	Ala	Gly	Leu	Arg	Ile	Thr	Ala	Pro	Leu	Met	Pro	Leu	Gly			
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Arg	Arg	Leu	Leu	Arg	Asn	Leu	Thr	Ala	Leu	Arg	Ile	Arg	Ala	Thr	Tyr			
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gga	gaa	tac	agt	act	ggg	tac	att	gac	aat	gtg	acc	ctg	att	tca	gcc	1221		

3/25

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Lys	Arg	Asp	Ser	Ala	Arg	Leu	Gly	Pro	Phe	Gly	Thr	Cys	Ile	Pro	Cys	
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Asn	Cys	Gln	Gly	Gly	Gly	Ala	Cys	Asp	Pro	Asp	Thr	Gly	Asp	Cys	Tyr	
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Ser	Gly	Asp	Glu	Asn	Pro	Asp	Ile	Glu	Cys	Ala	Asp	Cys	Pro	Ile	Gly	
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Phe	Tyr	Asn	Asp	Pro	His	Asp	Pro	Arg	Ser	Cys	Lys	Pro	Cys	Pro	Cys	
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His	Asn	Gly	Phe	Ser	Cys	Ser	Val	Ile	Pro	Glu	Thr	Glu	Glu	Val	Val	
465					470				475						480	
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Cys	Asn	Asn	Cys	Pro	Pro	Gly	Val	Thr	Gly	Ala	Arg	Cys	Glu	Leu	Cys	
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Ala	Asp	Gly	Tyr	Phe	Gly	Asp	Pro	Phe	Gly	Glu	His	Gly	Pro	Val	Arg	
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Pro	Leu	Ala	Pro	Asn	Pro	Ala	Asp	Lys	Cys	Arg	Ala	Cys	Asn	Cys	Asn	
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Pro	Met	Gly	Ser	Glu	Pro	Val	Gly	Cys	Arg	Ser	Asp	Gly	Thr	Cys	Val	
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Cys	Lys	Pro	Gly	Phe	Gly	Gly	Pro	Asn	Cys	Glu	His	Gly	Ala	Phe	Ser	
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4/25

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Lys Asn Leu Gly Asn Trp Lys Glu Glu Ala Gln Gln Leu Leu Gln Asn	
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Gly Lys Ser Gly Arg Glu Lys Ser Asp Gln Leu Leu Ser Arg Ala Asn	
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Leu Ala Lys Ser Arg Ala Gln Glu Ala Leu Ser Met Gly Asn Ala Thr	
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Phe Tyr Glu Val Glu Ser Ile Leu Lys Asn Leu Arg Glu Phe Asp Leu	
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Gln Val Asp Asn Arg Lys Ala Glu Ala Glu Glu Ala Met Lys Arg Leu	
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Ser Tyr Ile Ser Gln Lys Val Ser Asp Ala Ser Asp Lys Thr Gln Gln	
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Val Asp Thr Arg Ala Lys Asn Ala Gly Val Thr Ile Gln Asp Thr	
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Leu Asn Thr Leu Asp Gly Leu Leu His Leu Met Asp Gln Pro Leu	

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Arg Ala Lys Thr Gln Ile Asn	Ser Gln Leu Arg Pro	Met Met Ser	
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Glu Leu Glu Glu Arg Ala Arg	Gln Gln Arg Gly His	Leu His Leu	
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ctg gag aca agc ata gat ggg	att ctg gct gat gtg	aag aac ttg	3636
Leu Glu Thr Ser Ile Asp Gly	Ile Leu Ala Asp Val	Lys Asn Leu	
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gag aac att agg gac aac ctg	ccc cca ggc tgc tac	aat acc cag	3681
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Asp Asn Ser Gly Arg Cys Ser Cys Lys Pro Gly Val Thr Gly Ala Arg
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Cys Asp Arg Cys Leu Pro Gly Phe His Met Leu Thr Asp Ala Gly Cys
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Val Thr Gly Glu Arg Cys Asp Arg Cys Arg Ser Gly Tyr Tyr Asn Leu
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8/25

Asp Gly Gly Asn Pro Glu Gly Cys Thr Gln Cys Phe Cys Tyr Gly His
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Ser Ala Ser Cys Arg Ser Ser Ala Glu Tyr Ser Val His Lys Ile Thr
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Ser Thr Phe His Gln Asp Val Asp Gly Trp Lys Ala Val Gln Arg Asn
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Gly Ser Pro Ala Lys Leu Gln Trp Ser Gln Arg His Gln Asp Val Phe
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Ser Ser Ala Gln Arg Leu Asp Pro Val Tyr Phe Val Ala Pro Ala Lys
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Phe Leu Gly Asn Gln Gln Val Ser Tyr Gly Gln Ser Leu Ser Phe Asp
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Tyr Arg Val Asp Arg Gly Gly Arg His Pro Ser Ala His Asp Val Ile
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Glu His Pro Ser Asn Asn Trp Ser Pro Gln Leu Ser Tyr Phe Glu Tyr
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Asn Cys Gln Gly Gly Gly Ala Cys Asp Pro Asp Thr Gly Asp Cys Tyr

9/25

420

425

430

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10/25

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675 680 685

Asn Ser Tyr Gln Ser Arg Leu Asp Asp Leu Lys Met Thr Val Glu Arg
690 695 700

Val Arg Ala Leu Gly Ser Gln Tyr Gln Asn Arg Val Arg Asp Thr His
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Gly Phe Lys Ser Leu Ala Gln Glu Ala Thr Arg Leu Ala Glu Ser His
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785 790 795 800

Val Gly Ser Gly Ser Gly Ser Pro Asp Gly Ala Val Val Gln Gly Leu
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Glu Ala Thr Gln Ala Glu Ile Glu Ala Asp Arg Ser Tyr Gln His Ser
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Ser Phe Gln Val Glu Glu Ala Lys Arg Ile Lys Gln Lys Ala Asp Ser
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11/25

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12/25

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 atg cct gcg ctc tgg ctg ggc tgc tgc ctc tgc ttc tcg ctc ctc ctg 165
 Met Pro Ala Leu Trp Leu Gly Cys Cys Leu Cys Phe Ser Leu Leu Leu
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 aag tcc agg cag tgt atc ttt gat cgg gaa ctt cac aga caa act ggt 261
 Lys Ser Arg Gln Cys Ile Phe Asp Arg Glu Leu His Arg Gln Thr Gly
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 Cys Glu Lys Cys Lys Asn Gly Phe Tyr Arg His Arg Glu Arg Asp Arg
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 tgt ttg ccc tgc aat tgt aac tcc aaa ggt tct ctt agt gct cga tgt 405
 Cys Leu Pro Cys Asn Cys Asn Ser Lys Gly Ser Leu Ser Ala Arg Cys
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gac aac tct gga cgg tgc agc tgt aaa cca ggt gtg aca gga gcc aga Asp Asn Ser Gly Arg Cys Ser Cys Lys Pro Gly Val Thr Gly Ala Arg 100 105 110	453
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acc caa gac cag aga ctg cta gac tcc aag tgt gac tgt gac cca gct Thr Gln Asp Gln Arg Leu Leu Asp Ser Lys Cys Asp Cys Asp Pro Ala 130 135 140	549
ggc atc gca ggg ccc tgt gac gcg ggc cgc tgt gtc tgc aag cca gct Gly Ile Ala Gly Pro Cys Asp Ala Gly Arg Cys Val Cys Lys Pro Ala 145 150 155 160	597
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14/25

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15/25

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Cys	Pro	Ala	Cys	Tyr	Asn	Gln	Val	Lys	Ile	Gln	Met	Asp	Gln	Phe	Met	
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cag	cag	ctt	cag	aga	atg	gag	gcc	ctg	att	tca	aag	gct	cag	ggg	ggg	2037
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Glu	Gln	Ala	Leu	Gln	Asp	Ile	Leu	Arg	Asp	Ala	Gln	Ile	Ser	Glu	Gly	
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gct	agc	aga	tcc	ctt	ggg	ctc	cag	ttg	gcc	aag	gtg	agg	agc	caa	gag	2181
Ala	Ser	Arg	Ser	Leu	Gly	Leu	Gln	Leu	Ala	Lys	Val	Arg	Ser	Gln	Glu	
		675					680					685				
aac	agc	tac	cag	agc	cgc	ctg	gat	gac	ctc	aag	atg	act	gtg	gaa	aga	2229
Asn	Ser	Tyr	Gln	Ser	Arg	Leu	Asp	Asp	Leu	Lys	Met	Thr	Val	Glu	Arg	
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Val	Arg	Ala	Leu	Gly	Ser	Gln	Tyr	Gln	Asn	Arg	Val	Arg	Asp	Thr	His	
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Arg	Leu	Ile	Thr	Gln	Met	Gln	Leu	Ser	Leu	Ala	Glu	Ser	Glu	Ala	Ser	
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Gly	Phe	Lys	Ser	Leu	Ala	Gln	Glu	Ala	Thr	Arg	Leu	Ala	Glu	Ser	His	
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Val	Glu	Ser	Ala	Ser	Asn	Met	Glu	Gln	Leu	Thr	Arg	Glu	Thr	Glu	Asp	
		770			775						780					
tat	tcc	aaa	caa	gcc	ctc	tca	ctg	gtg	cgc	aag	gcc	ctg	cat	gaa	gga	2517
Tyr	Ser	Lys	Gln	Ala	Leu	Ser	Leu	Val	Arg	Lys	Ala	Leu	His	Glu	Gly	
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gtc	gga	agc	gga	agc	ggg	agc	ccg	gac	ggg	gct	gtg	gtg	caa	ggg	ctt	2565
Val	Gly	Ser	Gly	Ser	Gly	Ser	Pro	Asp	Gly	Ala	Val	Val	Gln	Gly	Leu	
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gtg	gaa	aaa	ttg	gag	aaa	acc	aag	tcc	ctg	gcc	cag	cag	ttg	aca	agg	2613
Val	Glu	Lys	Leu	Glu	Lys	Thr	Lys	Ser	Leu	Ala	Gln	Gln	Leu	Thr	Arg	
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gag	gcc	act	caa	gcg	gaa	att	gaa	gca	gat	agg	tct	tat	cag	cac	agt	2661
Glu	Ala	Thr	Gln	Ala	Glu	Ile	Glu	Ala	Asp	Arg	Ser	Tyr	Gln	His	Ser	
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18/25

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Cys Asp Arg Cys Leu Pro Gly Phe His Met Leu Thr Asp Ala Gly Cys
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Val Thr Gly Glu Arg Cys Asp Arg Cys Arg Ser Gly Tyr Tyr Asn Leu
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Asp Gly Gly Asn Pro Glu Gly Cys Thr Gln Cys Phe Cys Tyr Gly His
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Ser Ala Ser Cys Arg Ser Ser Ala Glu Tyr Ser Val His Lys Ile Thr
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Ser Thr Phe His Gln Asp Val Asp Gly Trp Lys Ala Val Gln Arg Asn
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Gly Ser Pro Ala Lys Leu Gln Trp Ser Gln Arg His Gln Asp Val Phe
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Ser Ser Ala Gln Arg Leu Asp Pro Val Tyr Phe Val Ala Pro Ala Lys
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Tyr Arg Val Asp Arg Gly Gly Arg His Pro Ser Ala His Asp Val Ile
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Pro Val Gly Tyr Lys Gly Gln Phe Cys Gln Asp Cys Ala Ser Gly Tyr
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660 665 670

Ala Ser Arg Ser Leu Gly Leu Gln Leu Ala Lys Val Arg Ser Gln Glu
675 680 685

Asn Ser Tyr Gln Ser Arg Leu Asp Asp Leu Lys Met Thr Val Glu Arg
690 695 700

Val Arg Ala Leu Gly Ser Gln Tyr Gln Asn Arg Val Arg Asp Thr His
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Arg Leu Ile Thr Gln Met Gln Leu Ser Leu Ala Glu Ser Glu Ala Ser
725 730 735

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Gly Phe Lys Ser Leu Ala Gln Glu Ala Thr Arg Leu Ala Glu Ser His
755 760 765

Val Glu Ser Ala Ser Asn Met Glu Gln Leu Thr Arg Glu Thr Glu Asp
770 775 780

Tyr Ser Lys Gln Ala Leu Ser Leu Val Arg Lys Ala Leu His Glu Gly
785 790 795 800

Val Gly Ser Gly Ser Gly Ser Pro Asp Gly Ala Val Val Gln Gly Leu
805 810 815

Val Glu Lys Leu Glu Lys Thr Lys Ser Leu Ala Gln Gln Leu Thr Arg
820 825 830

Glu Ala Thr Gln Ala Glu Ile Glu Ala Asp Arg Ser Tyr Gln His Ser

835

840

845

Leu Arg Leu Leu Asp Ser Val Ser Pro Leu Gln Gly Val Ser Asp Gln
 850 855 860

Ser Phe Gln Val Glu Glu Ala Lys Arg Ile Lys Gln Lys Ala Asp Ser
 865 870 875 880

Leu Ser Ser Leu Val Thr Arg His Met Asp Glu Phe Lys Arg Thr Gln
 885 890 895

Lys Asn Leu Gly Asn Trp Lys Glu Glu Ala Gln Gln Leu Leu Gln Asn
 900 905 910

Gly Lys Ser Gly Arg Glu Lys Ser Asp Gln Leu Leu Ser Arg Ala Asn
 915 920 925

Leu Ala Lys Ser Arg Ala Gln Glu Ala Leu Ser Met Gly Asn Ala Thr
 930 935 940

Phe Tyr Glu Val Glu Ser Ile Leu Lys Asn Leu Arg Glu Phe Asp Leu
 945 950 955 960

Gln Val Asp Asn Arg Lys Ala Glu Ala Glu Glu Ala Met Lys Arg Leu
 965 970 975

Ser Tyr Ile Ser Gln Lys Val Ser Asp Ala Ser Asp Lys Thr Gln Gln
 980 985 990

Ala Glu Arg Ala Leu Gly Ser Ala Ala Ala Asp Ala Gln Arg Ala Lys
 995 1000 1005

Asn Gly Ala Gly Glu Ala Leu Glu Ile Ser Ser Glu Ile Glu Gln
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Glu Ile Gly Ser Leu Asn Leu Glu Ala Asn Val Thr Ala Asp Gly
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Ala Leu Ala Met Glu Lys Gly Leu Ala Ser Leu Lys Ser Glu Met
 1040 1045 1050

Arg Glu Val Glu Gly Glu Leu Glu Arg Lys Glu Leu Glu Phe Asp
 1055 1060 1065

Thr Asn Met Asp Ala Val Gln Met Val Ile Thr Glu Ala Gln Lys
 1070 1075 1080

22/25

Val Asp Thr Arg Ala Lys Asn Ala Gly Val Thr Ile Gln Asp Thr
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gttattcagg ggatgagaat cctgacattg agtgtgctga ctgcccaatt ggtttctaca 180
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cccgtgtgta gctctgtgct gatggctact ttggggaccc ctttggtgaa catggcccag 360
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Leu Gly Pro Phe Gly Thr Cys Ile Pro Cys Asn Cys Gln Gly Gly Gly
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Asp Ile Glu Cys Ala Asp Cys Pro Ile Gly Phe Tyr Asn Asp Pro His

50

55

60

Asp Pro Arg Ser Cys Lys Pro Cys Pro Cys His Asn Gly Phe Ser Cys
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Ser Val Met Pro Glu Thr Glu Glu Val Val Cys Asn Asn Cys Pro Pro
85 90 95

Gly Val Thr Gly Ala Arg Cys Glu Leu Cys Ala Asp Gly Tyr Phe Gly
100 105 110

Asp Pro Phe Gly Glu His Gly Pro Val Arg Pro Cys Gln Pro Cys Gln
115 120 125

Cys Asn Asn Asn Val Asp Pro Ser Ala Ser Gly Asn Cys Asp Arg Leu
130 135 140

Thr Gly Arg Cys Leu Lys Cys Ile His Asn Thr Ala Gly Ile Tyr Cys
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Asp Gln Cys Lys Ala Gly Tyr Phe Gly Asp Pro Leu Ala Pro Asn Pro
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tgtgtgact gcccaattgg tttctacaac gatccgcacg accccgcag ctgcaagcca	240
tgtccctgtc ataacgggtt cagctgctca gtgattccgg agacggagga ggtggtgtgc	300
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ggggaccctt ttggtgaaca tggcccagtg aggcttgtc agccctgtca atgcaacagc	420
aatgtggacc ccagtgcctc tgggaattgt gaccggctga caggcaggtg tttgaagtgt	480
atccacaaca cagccggcat ctactgcgac cagtgcaaag caggctactt cggggaccca	540

ttggctccca acccagcaga caagtgtcga gcttgcaact gtaaccccat gggctcagag 600
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Cys Ile Cys Pro Val Gly Tyr Lys Gly Gln Phe Cys Gln Asp Cys Ala
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Ile Pro Cys Asn Cys Gln Gly Gly Gly Ala Cys Asp Pro Asp Thr Gly
 35 40 45

Asp Cys Tyr Ser Gly Asp Glu Asn Pro Asp Ile Glu Cys Ala Asp Cys
 50 55 60

Pro Ile Gly Phe Tyr Asn Asp Pro His Asp Pro Arg Ser Cys Lys Pro
 65 70 75 80

Cys Pro Cys His Asn Gly Phe Ser Cys Ser Val Met Pro Glu Thr Glu
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Glu Val Val Cys Asn Asn Cys Pro Pro Gly Val Thr Gly Ala Arg Cys
 100 105 110

Glu Leu Cys Ala Asp Gly Tyr Phe Gly Asp Pro Phe Gly Glu His Gly
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Pro Val Arg Pro Cys Gln Pro Cys Gln Cys Asn Asn Asn Val Asp Pro
 130 135 140

Ser Ala Ser Gly Asn Cys Asp Arg Leu Thr Gly Arg Cys Leu Lys Cys
 145 150 155 160

Ile His Asn Thr Ala Gly Ile Tyr Cys Asp Gln Cys Lys Ala Gly Tyr
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Phe Gly Asp Pro Leu Ala Pro Asn Pro Ala Asp Lys Cys Arg Ala Cys
180 185 190

Asn Cys Asn Pro Met Gly Ser Glu Pro Val Gly Cys Arg Ser Asp Gly
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Ala Phe Ser
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Gly Cys Arg Ser Asp Gly Thr Cys Val Cys Lys Pro Gly Phe Gly Gly
20 25 30

Pro Asn Cys Glu His Gly Ala Phe Ser
35 40

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(54) Title: USE OF ANTIBODIES TO THE GAMMA 2 CHAIN OF LALLMININ 5 TO INHIBIT TUMOR GROWTH AND METASTASIS

(57) Abstract: The present invention provides a methods and compositions for inhibiting tumor growth and/or metastasis involving the administering to a subject with a laminin 5-secreting tumor of an amount effective to inhibit tumor growth and/or metastasis of an antibody that binds to one or more epitopes of the laminin 5 γ 2 chain.

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INTERNATIONAL SEARCH REPORT

Application No
PCT/EP 03/12012

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According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 A61K C07K G01N C12Q C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, BIOSIS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2002/052307 A1 (KALLUNKI PEKKA ET AL) 2 May 2002 (2002-05-02) see inter alia (0091) and (0106) ---	1-13
X	US 5 660 982 A (PYKE CHARLES ET AL) 26 August 1997 (1997-08-26) claims ---	1-13
X	WO 02 30465 A (DELEU LAURENT ; LAND HARTMUT (US); UNIV ROCHESTER (US)) 18 April 2002 (2002-04-18) the whole document --- -/--	1-13

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

10 March 2004

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INTERNATIONAL SEARCH REPORT

Inventor's Application No
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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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X	<p>--- K. HELLMAN ET AL: "Cancer of the vagina: Laminin-5gamma2 chain expression and prognosis"</p> <p>INT J GYNECOL CANCER, vol. 10, 2000, pages 391-396, XP002272940 the whole document</p>	1-13
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INTERNATIONAL SEARCH REPORT

Information on patent family members

Application No

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